

Exercise and Brain Neurotransmission

Romain Meeusen and Kenny De Meirleir

Vrije Universiteit Brussel, Dept Human Physiology and Sportsmedicine, Laarbeeklaan,
Brussels, Belgium

Contents

Summary	160
1. Biosynthesis of Monoamines	161
1.1 The Dopaminergic System	161
1.2 The Noradrenergic System	162
1.3 The Serotonergic System	163
2. Exercise and Brain Monoamines	163
2.1 The Noradrenergic System	163
2.2 The Dopaminergic System	166
2.3 The Serotonergic System	170
2.4 Neurotransmitter Interactions	174
3. Neurotransmission and Exercise Performance	175
3.1 Tryptophan, Branched Chain Amino Acids and Neurotransmission	175
3.2 Precursor Loading, Pharmacological Manipulation and Neurotransmission	179
4. Measurement of Extracellular Neurotransmitter Levels	181
5. Conclusions	185

Summary

Physical exercise influences the central dopaminergic, noradrenergic and serotonergic systems. A number of studies have examined brain noradrenaline (norepinephrine), serotonin (5-hydroxytryptamine; 5-HT) and dopamine with exercise. Although there are great discrepancies in experimental protocols, the results indicate that there is evidence in favour of changes in synthesis and metabolism of monoamines during exercise.

There is a possibility that the interactions between brain neurotransmitters and their specific receptors could play a role in the onset of fatigue during prolonged exercise. The data on the effects of branched chain amino acid (BCAA) supplementation and 'central fatigue' seem to be conflicting, although recent studies suggest that BCAA supplementation has no influence on endurance performance.

There are numerous levels at which central neurotransmitters can affect motor behaviour; from sensory perception, and sensory-motor integration, to motor effector mechanisms. However, the crucial point is whether or not the changes in neurotransmitter levels trigger or reflect changes in monoamine release. Until recently most studies were done on homogenised tissue, which gives no indication of the dynamic release of neurotransmitters in the extracellular space of living organisms.

Recently, new techniques such as microdialysis and voltammetry were introduced to measure *in vivo* release of neurotransmitters. Microdialysis can collect virtually any substance from the brain of a freely moving animal with a limited

amount of tissue trauma. This method allows measurement of local neurotransmitter release during on-going behavioural changes such as exercise.

The results of the first studies using these methods indicate that the release of most neurotransmitters is influenced by exercise. Although the few studies that have been published to date show some discrepancies, we feel that these recently developed and more sophisticated *in vivo* methods will improve our insight into the relationship between the monoamine and other transmitters during exercise. Continued quantitative and qualitative research needs to be conducted so that a further understanding of the effects of exercise on brain neurotransmission can be gained.

The health benefits of exercise include favourable physiological, psychological and biochemical changes.^[1] Research into the physiological effects of exercise is usually on the muscular or neuromuscular systems even though it is apparent that there is also an influence on the CNS, with convincing evidence that several neurotransmitters are involved in control of locomotion.^[2-8]

Fatigue during prolonged exercise has traditionally been attributed to the occurrence of a 'metabolic end-point', where muscle glycogen levels are depleted, plasma glucose levels are reduced, and plasma free fatty acid levels are elevated.^[9] There is also a 'central fatigue hypothesis'^[10-12] which is based on the increase in the level of brain serotonin (5-hydroxytryptamine; 5-HT) during exercise. However, the physiological mechanisms for central fatigue are largely unexplored.

This review will focus on the effects of exercise on neurotransmission, especially the influence of exercise on the monoaminergic systems. We will discuss the possible role and interaction of the neurotransmitters and their precursors in central fatigue and also in motor behaviour. Finally, recent developments in the direct measurement of neurotransmitter release with microdialysis and voltammetry are presented.

This work includes both animal and human studies. Most of the studies that examined the effects of exercise on brain neurotransmitters were performed on animals. When exercise performance and precursor loading or pharmacological manipulations are discussed we also included the results of human studies.

1. Biosynthesis of Monoamines

The biogenic amines include the catecholamines, dopamine, noradrenaline (norepinephrine), adrenaline (epinephrine), and the indolamine, serotonin. Tyrosine is the common amino acid precursor of all catecholamines, while the precursor of serotonin is the essential amino acid tryptophan.

Monoaminergic neurons modulate a wide range of functions in the central nervous system.^[13] Noradrenergic neurons are involved in cardiovascular function, sleep and analgesic responses, while dopaminergic neurons are linked with motor function^[2] and serotonergic activity is associated with pain, fatigue, appetite and sleep.^[13]

1.1. The Dopaminergic System

Dopaminergic cell groups are found in the mesencephalon, the diencephalon and the telencephalon. The main ascending dopaminergic pathways include the nigrostriatal tractus, the ventral mesostriatal (or mesolimbic) pathway and the tubero-infundibular system which arises from cells located in the diencephalon.^[14]

The rate-limiting step in the biosynthesis of dopamine is the hydroxylation of tyrosine to dihydroxyphenylalanine (dopa) by the enzyme tyrosine hydroxylase. The majority of tyrosine hydroxylase is located in catecholamine nerve terminals. Tyrosine hydroxylase activity can be inhibited by the catecholamines, suggesting a feedback inhibitory effect. Dopa is decarboxylated to dopamine by the enzyme dopa-decarboxylase (aromatic amino acid decarboxylase). The activity of

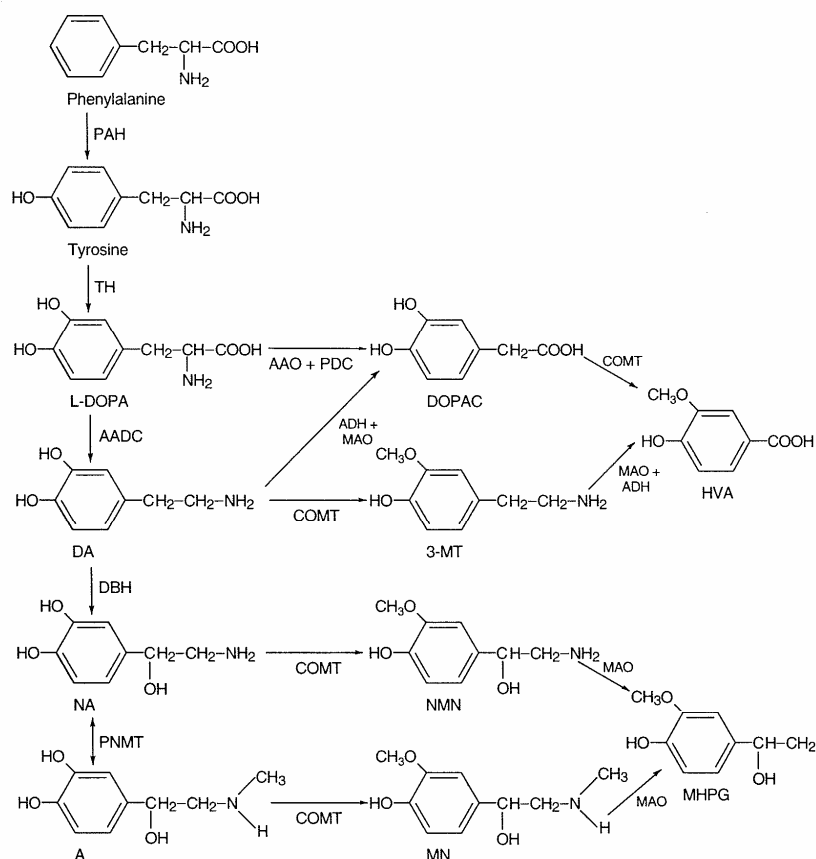


Fig. 1. Biosynthesis and catabolism of the catecholamines. *Abbreviations:* A = adrenaline (epinephrine); AADC = aromatic L-amino acid decarboxylase; AAO = L-amino acid oxidase; ADH = alcohol dehydrogenase; COMT = catechol-O-methyl transferase; DA = dopamine; DBH = dopamine β -hydroxylase; DOPAC = 3,4-dihydroxyphenylacetic acid; HVA = homovanillic acid; L-dopa = L-dihydroxyphenylalanine; MAO = monoamine oxidase; MHPG = 3-methoxy-4-hydroxyphenylethylene glycol; MN = metanephrine; 3-MT = 3-methoxytyramine; NA = noradrenaline (norepinephrine); NMN = normethanephrine; TH = tyrosine hydroxylase; PAH = phenylalanine hydroxylase; PDC = pyruvate decarboxylase; PNMT = phenylalanine-N-methyltransferase.

this enzyme is not rate-limiting in the synthesis of the catecholamines, and is therefore no regulating factor in their formation. Dopamine is in normal physiological conditions first metabolised to 3,4-dihydroxyphenylacetic acid (DOPAC) by monoamine oxidase and aldehyde oxidase. DOPAC is then further metabolised into homovanillic acid by catechol-O-methyltransferase.^[14]

1.2 The Noradrenergic System

The neurons that synthesise noradrenaline (nor-epinephrine) are restricted to the pontine and medullary tegmental region. The locus coeruleus is quantitatively the most important noradrenergic nucleus in the brain. Its efferent fibres constitute a major ascending pathway, the dorsal noradrenergic

bundle. Along its course different branches emerge to innervate a large number of mesencephalic areas (dorsal raphe nucleus, thalamus, hypothalamus, hippocampus, septum and cortex).

In the noradrenergic neurons dopamine is converted into noradrenaline (norepinephrine) through dopamine β -hydroxylase.^[15] The enzymes responsible for the catabolism of noradrenaline are monoamine oxidase and catechol-*O*-methyltransferase. The main metabolite of noradrenaline is 3-methoxy-4-hydroxyphenylethyleneglycol (MHPG) [fig. 1].

1.3 The Serotonergic System

Serotonin-containing neurons are present in the mesencephalon, pons and medulla oblongata. They are mainly located in the raphe nuclei. Efferent fibres innervate the substantia nigra, various thalamic centres, the nucleus caudatus, the putamen, the nucleus accumbens, the cortex, and the hippocampus. Other serotonergic cells innervate the ventral horn of the spinal cord and the medulla.^[14]

The synthesis of serotonin requires two enzymatic steps. The dietary amino acid precursor tryptophan is first hydroxylated by a tryptophan hydroxylase to L-5-hydroxytryptophan (5-HTP) and then decarboxylated to serotonin. Serotonin itself is metabolised by 2 enzymes (aldehyde dehydro-

genase and monoamine oxidase) to 5-hydroxyindoleacetic acid (5-HIAA)[fig. 2].

2. Exercise and Brain Monoamines

The first reports that examined the influence of exercise on brain neurotransmitters appeared in the 1960s.^[16-19] These studies mostly used exercise as a stress model, or compared exercise with other stressors such as exposure to cold.^[17] Since then several studies have continued to use this approach comparing or combining exercise with other stressors such as foot-shock,^[20] cold,^[17] tail pinch, immobilisation, or restraint.^[21,22] Other studies described the influence of exercise on brain monoamines as a possible intervention in affective disorders^[23,24] and depression.^[25-27]

Most of these animal studies, however, examined brain monoamine levels with acute and chronic exercise protocols to explore the effects of a physiological stimulus on brain neurotransmission.

2.1 The Noradrenergic System

Studies that examined whole brain noradrenaline levels after acute bouts of exercise (running or swimming) mostly found a decrease,^[16,18,28] no effect,^[19,29] or a small not-significant increase in

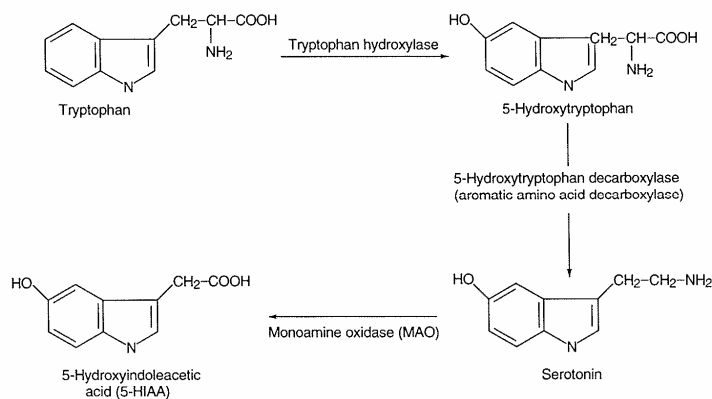


Fig. 2. Biosynthesis and catabolism of serotonin (5-hydroxytryptamine; 5-HT).

Table I. The effects of exercise on noradrenaline (norepinephrine)

Reference	Animals	Exercise	Training	Brain area	Results																
Cicardo et al. ^[28]	Wistar rats Male n = 60	Swimming in 23°C	To exhaustion	Whole brain	↓																
Moore ^[19]	Albino mice Male n = 8	Running wheel	Spontaneous activity 10 mins	Whole brain	Administration of αMT: NA ↓																
Moore & Larivière ^[18]	S-D rats Female n = 84 (7 groups)	Swimming	4h 23°C or 37°C	Whole brain	↓																
Barchas & Freedman ^[16]	S-D Male (200g) n = 50 each group	Treadwheel: 3h, 1.8 m/min Swim to exhaustion	15°C: 15 → 30 mins 23°C: 4 → 6h	Whole brain	10% ↓ 11% ↓ 26% ↓																
Sheldon et al. ^[29]	Swiss-Webster mice Female (25-30g)	Treadmill: 5.4 m/min	150 mins + tyrosine C ¹⁴	Whole brain	No change in synthesis and turnover of catecholamines (no specific determination)																
Broocks et al. ^[40]	Wistar rats Male n = 168 (5 groups)	Running wheel	Spontaneous running 1 group + food deprivation	Hypothalamus Preoptic area	↑ In both groups ↑ In both groups																
Rea & Hellhammer ^[36]	Wistar rats n = 21	Running wheel	Spontaneous running	Pons medulla Thalamus Septum Neocortex Cerebellum Midbrain Hypothalamus Striatum Hippocampus	<table style="border: none;"> <tr> <td style="text-align: center;">NA</td> <td style="text-align: center;">MHPG</td> </tr> <tr> <td style="text-align: center;">~</td> <td style="text-align: center;">↑ In all runner groups</td> </tr> <tr> <td style="text-align: center;">~</td> <td style="text-align: center;">} compared to control except</td> </tr> <tr> <td style="text-align: center;">~</td> <td style="text-align: center;">for septum, striatum, hippocampus</td> </tr> <tr> <td style="text-align: center;">~</td> <td style="text-align: center;">↑ In sedentary food restricted compared to control</td> </tr> <tr> <td style="text-align: center;">~</td> <td style="text-align: center;">↑ In running compared to control, food restricted</td> </tr> <tr> <td style="text-align: center;">~</td> <td style="text-align: center;">↑ In runners - control</td> </tr> <tr> <td style="text-align: center;">~</td> <td style="text-align: center;">↓ In runners - control</td> </tr> </table>	NA	MHPG	~	↑ In all runner groups	~	} compared to control except	~	for septum, striatum, hippocampus	~	↑ In sedentary food restricted compared to control	~	↑ In running compared to control, food restricted	~	↑ In runners - control	~	↓ In runners - control
NA	MHPG																				
~	↑ In all runner groups																				
~	} compared to control except																				
~	for septum, striatum, hippocampus																				
~	↑ In sedentary food restricted compared to control																				
~	↑ In running compared to control, food restricted																				
~	↑ In runners - control																				
~	↓ In runners - control																				
Stone ^[33]	S-D Male n = 22	Motor driven running wheel	3h 5.5 m/min → 8 m/min	Hypothalamus Brainstem	NA ↓, MHPG ↑; reserpine: NA ↓, ↑ MHPG αMT: NA ↓ -, MHPG ↓																
Östman & Nyback ^[32]	S-D Male n = 24	Swimming in 35°C	17 weeks, 1 → 2.5 h/day	Whole brain	↑																
Acworth et al. ^[12]	Male rats n = 23 (± 5/group)	Treadmill: 25 m/min 4° incline Treadmill 90 mins: 25 m/min 4° incline	90 mins 5 weeks, 6 days/week 60 mins/day incline 1° → 4°	Whole brain Whole brain	~ 21% ↑ sig.																
Brown & Van Huss ^[30]	S-D Male n = 80	Running wheel: 60 mins	8 weeks, 1 h/day	Whole brain	↑ In trained animals αMT depleted brain NA																
Decastro & Duncan ^[25]	Long-Evans hooded rats Male (290-370g) n = 12	Running wheel: Animals killed 48h after final training	Operant conditioning: 8 weeks, 5 days/week, 2 h/day (ran ± 1.2 km/day); not conditioned animals ran ± 0.5 km/day	Whole brain	Small ↑ (NS) ³H-piperone binding (NS) ↓																

Table I. Contd

Reference	Animals	Exercise	Training	Brain area	Results
Brown et al. ^[31]	Wistar Female n = 40	Treadmill 30 m/min	8 weeks, 5 days/week, 30 mins/day (normal diet or fat diet)	Whole brain	↑
				Cerebellum	↑
				Midbrain	↑
Brown et al. ^[41]	Female rats n = 36	Treadmill	30 mins, 6 weeks	Telencephalon (mainly Cortex)	Sedentary ↓ Trained =
				Medulla oblongata	↓ ↑
				Hypothalamus	↓ ↓
Elam et al. ^[23]	Wistar Koyoto Male n = 25	Running wheel: spontaneous running, animals killed immediately or 24h after last running period	5km/12h 7 days	Limbic forebrain, striatum, brain stem, cortex, spinal cord	↑ NA synthesis in brain stem
Blomstrand et al. ^[39]	Wistar rats Female (200-220g) n = ±5/group	Treadmill: sedentary, ±17m/min 7° incline RTE: trained, ±30-35 m/min 7° incline	Sedentary accommodation: 10 mins/day → low speed Trained: 11 weeks 6 days/week 1 h/day, 3 cm/min 7° incline	Cortex	Sedent. Not measured Trained Not measured
				Cerebellum	~ ~
				Hippocampus	~ ~
				Striatum	67% ↑ 55% ↑
				Brainstem	~ ~
Sudo ^[37]	S-D 7/group	Swimming 4h: 35°C water		Hypothalamus	Sig. ↓ from 15 mins → 4h
				Pons medulla	Sig. ↓ at 15 mins, 2h, 4h
				Midbrain	Sig. ↓ at 15 mins, 1h, 2h, 4h
Gordon et al. ^[17]	S-D Female (160-220g) n = 46 (TTL)	Rotating drum: 7 rpm (high speed)	3 and 5h (1 exc group with αMT) 1h + tyrosine C ¹⁴	Brain stem	↓ With exc (slight depletion) ↓↓ With αMT + exc ↑ Radioactivity with exc ↑ Synthesis of NA
Heyes et al. ^[35]	S-D Male (350-400g) n = 25 (± 5/group)	Treadmill: 36 m/min	RTE or after (6, 11, 16.5 mins)	Striatum	~
				Brainstem	↓ Progressively during exc
				Hypothalamus	
Lukaszyk et al. ^[38]	Wistar rats Male (150-220g) n = 5/group	Treadmill: 30 m/min	20 mins	Cortex, striatum	Sig. ↑
				Hypothalamus	Sig. ↓
				Midbrain,	=
				cerebellum	

Abbreviations: αMT = α-methyl-*p*-tyrosine; exc = exercise; h = hour(s); MHPG = 3-methoxy-4-hydroxyphenylethyleneglycol, NA = noradrenaline; NS = not significant; RTE = run to exhaustion; S-D = Sprague-Dawley rats; sig. = significant; = indicates no significant change; ↑ = increase; ↓ = decrease; ~ = no significant difference.

brain noradrenaline level.^[12,25] (See table I.) Whole brain noradrenaline levels increased after chronic exercise training.^[12,25,30-32]

Some studies examined noradrenaline levels in specific brain regions. Noradrenaline levels decreased due to acute exercise in brain stem,^[17,33-35] hippocampus,^[36] pons-medulla,^[37] midbrain,^[37] and hypothalamus,^[33,35,37,38] while noradrenaline level in the striatum,^[36,39] cortex^[38] and preoptic area^[40] increased. Interestingly, hypothalamic

noradrenaline levels have been shown to increase in food-restricted rats.^[36] The same study measured MHPG and found an increase in most brain regions indicating an activation of noradrenaline catabolism.

Studies that examined the effects of exercise training on noradrenaline levels in different brain regions found mostly an increase or no significant result.^[30,31,39,41] Stone^[33] examined alterations in storage of [³H]noradrenaline in hypothalamus of

rats during 3 hours of exercise on a motor driven running wheel. They further compared exercise with injections of reserpine (inhibition noradrenaline storage) or α -methyl-*p*-tyrosine (inhibition noradrenaline synthesis). Running did not alter storage of noradrenaline, therefore the authors concluded that it was likely that the noradrenaline depletion during running was derived from newly synthesised noradrenaline and not from reuptake mechanisms.^[33] One study^[23] investigated the accumulation of dopa as an index of tyrosine hydroxylation activity in order to obtain an indication of the monoamine synthesis rate. It found higher dopa levels in the brain stem indicating an increased synthesis of noradrenaline in this predominantly noradrenaline-rich region.^[23]

Only one study reported changes in adrenaline level following 4 hours of swimming.^[37] The adrenaline levels in hypothalamus, pons-medulla and midbrain showed a gradually decrease, with a significant decrease in the second part of the exercise period.^[37]

It seems that acute exercise results in a depletion of brain noradrenaline probably because of an acceleration in noradrenaline turnover by activating tyrosine hydroxylase activity,^[15] while chronic exercise has been found to elevate brain noradrenaline levels. These adaptations are region specific.

2.2 The Dopaminergic System

Dopamine neurons are considered to be critical components in the motor system.^[2] A number of studies have examined the effect of acute or chronic exercise on dopamine synthesis and metabolism (table II). Two studies^[17,29] used the incorporation of [¹⁴C]tyrosine into [¹⁴C]dopamine and did not find changes in central dopamine synthesis and turnover. Another early study,^[21] however, found an increase in homovanillic acid level in mice following swimming and running. Chaouloff et al.^[42] also confirmed an increased dopamine metabolism in the whole brain of rats from running.

A number of studies used trained animals to study the effects of chronic exercise or the effects of an acute exercise session following training on

brain dopamine levels. Whole brain dopamine level was increased in rats killed 48 hours after an 8-week training period.^[25] A 1-week training model was used to examine brain dopamine metabolism and found that the sum of the levels of DOPAC and homovanillic acid was increased with running and remained elevated throughout the first hour of recovery.^[43] Two studies^[12,30] did not find a significant influence of exercise on whole brain dopamine level of trained rats.

Most studies examined the effects of acute and chronic exercise on regional dopaminergic systems. One study^[38] examined the effects of 20 minutes of exercise on regional dopamine levels in untrained rats and found decreased levels in all brain regions examined. Swimming for 4 hours in 35°C water resulted in a small but not significant decrease in dopamine level in striatum and midbrain.^[37] Other studies found increased dopamine, DOPAC and homovanillic acid in striatum,^[20,35] brainstem^[35] and hypothalamus.^[39]

In the same region (the hypothalamus) the level of dopamine and its metabolites was unchanged although the ratio DOPAC + homovanillic acid to dopamine increased.^[35] These results indicate an increased dopamine synthesis and metabolism, although in another study,^[17] treatment with α -methyl-*p*-tyrosine had no effect on dopamine synthesis in brainstem, and it decreased DOPAC level in the whole brain in control and exercising rats.^[43]

The fact that dopamine metabolism in rat striatum is involved in movement has been shown.^[2] The data showed that there is a very close relationship between dopamine production and all aspects of motor behaviour (speed, direction and body posture). The dopamine level in the nucleus accumbens appears to be a marker for the speed of animals while dopamine level in the caudate is more related to posture.^[2]

The effect of training on regional dopamine level was examined in several studies. In some studies the animals ran, while others examined regional transmitter levels without an acute training session. The level of dopamine and its metabolites was found to increase in hypothalamus,^[9,41,43] mid-

Table II. The effects of exercise on dopamine

Reference	Animals	Exercise	Training	Brain area	Results
Sheldon et al. ^[26]	Swiss-Webster mice Female (25-30g) n = 6/group	Treadmill: 5.4 m/min, 150 mins + tyrosine C ¹⁴		Whole brain	No change in synthesis and turnover of catecholamines
Bliss & Allison ^[21]	Mice Male ± 35g	Swimming: 1h 37°C Spontaneously running: 30 mins		Whole brain	Swimming: ↑ HVA, ↑ DA metabolism Running: ↑ HVA
Freed & Yamamoto ^[23]	S-D Male (300-350g) n = 6/group	Treadmill: 20 mins 1.4, 2 → 7.2 m/min		Striatum: n. caudatus n. accumbens	↑ DA & DOPAC (posture & direction) ↑ DA & DOPAC (speed & direction)
Speciale et al. ^[20]	S-D Male (250-300g) n = 6/group	Running wheel: 6.3 m/min 1h 12.6 m/min		Striatum: n. caudatus, n. accumbens, prefrontal cortex	↑ DA metabolism in striatum not front cortex
Gordon et al. ^[17]	S-D Female (160-220g) n = 46 (TTL)	Rotating drum 7 rpm (high speed) 3 and 5h (1 exc. group with αMT) 1h + tyrosine C ¹⁴ group		Brainstem	No significant effect on DA synthesis
Heyes et al. ^[25]	S-D Male (350-400g) n = 25 (±5/group)	Treadmill: 36.0 m/min RTE or after 6, 11, 16.5 mins		Striatum Brainstem Hypothalamus	DA ↑, DOPAC ↑, HVA ↑, [DOPAC + HVA]/DA ↑ DA ↑, DOPAC ↑, HVA ↑, [DOPAC + HVA]/DA ↑ DA =, DOPAC =, HVA =, [DOPAC + HVA]/DA ↑ (late exhaustion rats)
Lukaszuk et al. ^[28]	Wistar rats Male (150-220g) n = 5/group	Treadmill: 20 mins 30 m/min		Cortex, hippocampus, hypothalamus, striatum, midbrain cerebellum	↓ DA in all regions NS in hippocampus
Acworth et al. ^[12]	Male rats	Treadmill: 90 mins 25 m/min 4° incline	5 weeks, 6 days/week, 60 mins/day incline 1° to 4°	Whole brain	-[DA]
Chaouloff et al. ^[42]	Wistar rats Male (250-300g) n = 5 (brain) n = 6 (CSF)	Treadmill: 60 mins, 20 m/min	1 week, 4 to 5 times, last session 1h at 20 m/min	Whole brain CSF withdrawal	HVA ↑, DOPAC ↑ indicating DA metabolism with exc ↑ HVA ↑, DOPAC ↑ NS
Chaouloff et al. ^[43]	Wistar rats Male (220-250g) n = 5/group	Treadmill: 22 m/min 60 mins	1 week, 4 to 5 times, last session 1h at 20 m/min	Whole brain	DOPAC + HVA ↑ remained sig. ↑ during first h recovery AMPH: DOPAC ↓ in control and exc AMPH + exc: DOPAC higher than control αMPT: DOPAC ↓ in control and exc Haloperidol (post exc): DOPAC ↑ in control and exc
Bailey et al. ^[44]	Wistar rats Female n = 5	Treadmill: 60 mins, 20 m/min, 5% incl.	Treadmill accommodation: 3-4 weeks speed + incl. ramp, 1 last session: 20 m/min 5% 30 mins	Striatum Frontal cortex Midbrain Hypothalamus Hippocampus	DA =, DOPAC ↑, HVA ↑ DA =, DOPAC =, HVA ↑ DA ↑, DOPAC ↑, HVA ↑ DA ↑, DOPAC ↑, HVA ↑ DA ↑
				Hippocampus Striatum Midbrain	Exc + mCPP DA ↑, DOPAC = DA =, DOPAC = DA ↑, DOPAC ↑ (but lower than exc alone)

Continued on next page

Reference	Animals	Exercise	Training	Brain area	Results	Trained
Blomstrand et al. ^[39]	Wistar rats Female (200-220g) n = ± 5/group	Treadmill: sedentary, ±17 m/min 7° incline RTE: trained, ±30-35 m/min 7° incline 24h after last running period	Sedentary accommodation 10 mins/day at slow speed Trained: 1 week 6 days/week 1 h/day	Cortex Cerebellum Hippocampus Striatum Brainstem Hypothalamus	Sedentary Not measured Below detect limit - 27% ↑ 61% ↑ (sig.) Immed. post run DOPA ↓ DOPA ↓ DOPA ↑	Trained Not measured Below detect limit - 65% ↑ NS 36% ↑ NS 24h post run = (decreased DA synthesis) =
Eliam et al. ^[23]	Wistar-koyoto Male n = 25	Running wheel: spontaneous running, animals killed immediately or 24h after last running period	5 km/12h 7 days	Limbic forebrain (incl. n. accumbens) Striatum Brainstem, cortex, spinal cord	DOPA ↓ DOPA ↓ DOPA ↑	
Hoffmann et al. ^[24]	Spontaneously hypertensive rats Male (200-330g) n = 32 (8/group)	Running wheel, spontaneous running, animals killed 1-2h, or 47-48h after last exc. period	5 week period: 4 km/24h TTL: 7 weeks	Limbic forebrain Cortex, striatum, diencephalon, brainstem	↓ DOPAC (immed. post run) DA turnover (DOPAC/DA); unaltered No effect in either period cMT did not sig. lower [DA]	
Brown & Van Huss ^[30]	S-D Male n = 80	Running wheel 60 mins	8 weeks, 1h/day	Whole brain		
Brown et al. ^[41]	Female rats n = 36	Treadmill 30 mins	8 weeks, 1h/day	Telencephalon (mainly cortex), midbrain, oblongata Hypothalamus	Sedentary ↑ ↓ ↓	Trained ↑ ↑ =
Decastro & Duncanson ^[25]	Long-Evans hooded rats Male (290-370g) n = 12 (6 pairs)	Running wheel: Animals killed 48h after final training	Operant conditioning technique 8 weeks, 5 days/week, 2h/day (ran ± 1.2 km/day) (not conditioned animals ran ± 0.5 km/day)	Whole brain	DA ↑ - ³ H-spiperone binding ↓ compared to non-reinforced animals but NS	
Gilliam et al. ^[45]	S-D Male (530-555g) n = 30	Treadmill: animals killed 48h after final training	Endurance, 27 m/min, 12 weeks, 6 days/week on interval training	Striatum	Endurance & interval group were not sig. different from each other - ³ H-spiperone binding sig. ↑ in runners	
MacRae et al. ^[46]	S-D Male young adults	Treadmill	6 months, 6 days/week, 27 m/min	Striatum	No effect on steady-state levels of [3H]spiperone binding DA binding sites exc can alter the number of DA binding sites	
Bailey et al. ^[9]	Wistar rats Male (300-400g) TTL = 72 (n = 8/group)	Treadmill: 60 mins or exhaustion, 20 m/min, 5° incline	Treadmill accommodation, 3-4 weeks, speed + run time ↑; last session: 20 m/min, 5%, 30 mins	Midbrain Striatum Hypothalamus Hippocampus	1h DA ↑ DOPAC ↑ DA ↑ DOPAC ↑ DA ↑ DOPAC ↑ DA = DOPAC = 1h OD DA ↓ DOPAC = DA = DOPAC = DA = DOPAC ↑ NS DA = DOPAC = 1h LY 53857 DA ↑ DOPAC ↑ DA ↑ DOPAC ↑ DA ↑ DOPAC ↑ DA = DOPAC =	Exh DA ↓ DOPAC ↑ DA ↑ DOPAC ↑ DA = DOPAC = DA = DOPAC ↑ NS EXH OD DA ↓ DOPAC = DA = DOPAC = DA = DOPAC = DA = DOPAC = EXH LY 53857 DA ↑ DOPAC ↑ DA ↑ DOPAC ↑ DA ↑ DOPAC ↑ DA = DOPAC =

Wilson & Marsden ^[66]	Lister hooded rats Male (170-230g) n = 12	Treadmill: 60 mins, 20 m/min	4 weeks accommodation Speed & run time gradually increased	Hypothalamus	1 DA
Sudo ^[67]	S-D 7/group	Swimming: 4h, 35°C water		Hypothalamus Pituitary Midbrain	Sig. ↓ from 15 mins → 4h Sig. ↓ at 15 mins, 2h, 4h Sig. ↓ at 15 mins, 1h, 2h, 4h

Abbreviations and symbols: αMT = α-methyl-p-tyrosine; AMPH = amphetamine; CSF = cerebrospinal fluid; DA = dopamine; DOPAC = 3,4-dihydroxyphenylacetic acid; exc = exercise; EXH = exhaustion; h = hour(s); LY 53857 = serotonin antagonist; mCPP = 5-HT_{1A} agonist; MHPG = 3-methoxy-4-hydroxyphenylethyl(meglycol); NS = not significant; QD = quipazine dimeclate (serotonin agonist); RTE = run to exhaustion; S-D = Sprague-Dawley rats; SEM = standard error; sig. = significant; TTL = total; ↑ = increase; ↓ = decrease; - = no significant difference.

brain,^[9,43,44] prefrontal cortex,^[43] hippocampus^[43,44] and striatum.^[9,41,43] Bailey et al.^[9] also found elevated dopamine and DOPAC levels after 1 hour of exercise, however, these increases were not as great at exhaustion. The hippocampal dopamine levels did not change as a result of exercise.^[9]

Another group^[23] used spontaneous, long term running in a wheel cage. Exercising animals were killed 1 to 2 hours or 24 hours after the last running period. By giving the animals NSD (an aromatic amino acid decarboxylase inhibitor) 30 minutes before killing the authors measured the accumulation of dopa, an index for monoamine synthesis rate in different brain regions. The results showed a decreased rate of dopamine synthesis in dopamine-rich brain regions, while dopa levels were considerably higher in noradrenaline regions (brainstem) indicating an increased synthesis of noradrenaline in this region. All these alterations were normalised after 24 hours. However, measurement of dopa accumulation does not enable an evaluation of the *in vivo* physiological activity of the monoamine pathways.^[23]

Three studies^[25,45,46] used [³H]spiperone receptor binding to get an indication of transmitter dynamics. An operant conditioning model with positive reinforcement has been used to induce exercise.^[25] Although dopamine levels increased, they found [³H]spiperone binding to be decreased in whole brain homogenates. The authors state that caution must be exercised in interpreting these results as a change solely in dopamine receptor binding. A more specific regional determination could result in different findings. This was confirmed by Gilliam et al.^[45] who showed that animals, exercised on a moderate to high intensity endurance or interval running protocol, showed significantly higher [³H]-spiperone receptor binding than sedentary controls.

The effects of 6 months of endurance training on the relationships among steady-state *in vivo* levels of dopamine and its metabolites and the affinity and density of striatal-D₂ receptors were determined.^[46] Endurance training had no effect on steady-state levels of dopamine or its major meta-

bolites in striatum. The major finding of this study was that 6 months of endurance training alters neurochemical markers in the nigrostriatal dopamine system in young adult rats. Binding of [³H]-spiperone to D₂ receptors and the ratio of binding to levels of DOPAC were enhanced in the runners. Together these results suggest that a shift in dopamine function may occur as a result of exercise, either due to altered dopamine release or to changes in D₂ binding sites.^[46] However, as has already been indicated, spiperone will also bind to other (serotonin) receptor types.^[15]

It is difficult to draw conclusions from these studies because there is no uniformity with the study methods used. Neurotransmitter levels in whole brain or brain regions are just an indication of the amount neurotransmitter, and give us no information concerning neuronal activity. Receptor binding studies used are not specific to one receptor type.

The dopaminergic nerve terminals appear to play an important role in the regulation of locomotor activity^[47] and it seems that the influence of acute or chronic exercise is region specific. However, it should be recognised that the so called 'motor circuit' containing neurons from the striatum, substantia nigra, cerebral cortex and thalamus, interact constantly through several transmitters and receptor types.^[48] It is difficult, if not impossible, to register this dynamic and constant interaction with brain homogenate preparations.

2.3 The Serotonergic System

A number of studies have examined brain serotonin and 5-HIAA levels with acute and chronic exercise (table III). Chalouff and his co-workers have published several papers on this topic.^[15,42,43,49-52] Except for one study^[28] that found no change in serotonin level, whole brain serotonin and 5-HIAA increase following an acute bout of exercise.^[12,16,53] However, in trained rats it seems that brain serotonin level is unaltered while 5-HIAA level increases. The first studies by Chalouff et al.^[42,49] showed that exercise increased brain and cerebrospinal fluid tryptophan and 5-

HIAA, indicating an increase in serotonin synthesis and metabolism. The same authors^[51] did not find a difference in basal brain serotonin levels between short and long term trained rats. A single running session did not change serotonin level in the rats trained for 1 week, but serotonin was diminished in rats trained for 8 weeks, probably indicating a different serotonin utilisation.^[51]

Acute and chronic exercise studies have found both increased and decreased levels and turnover of serotonin and 5-HIAA, depending on the brain region of interest. Striatal, hippocampal and mid-brain serotonin and 5-HIAA levels increased after a training session, or after an acute bout of exercise in trained rats.^[9,31,39,44,52,54]

Dey et al.^[26] studied serotonin and 5-HIAA alterations in different brain regions following acute (1 hour swim) and chronic exercise (4 weeks of swimming, 6 days/week). Acute exercise significantly increased the synthesis and metabolism of serotonin in the brain stem and hypothalamus, and there were no changes in cerebral cortex and hippocampus. Chronic exercise activated not only the synthesis but also the metabolism of serotonin in cerebral cortex. One week after the termination of training this neuronal adaptation was still present. In brain stem, serotonin turnover increased immediately after the training session. In hippocampus a delayed effect was observed, because serotonin level was unaltered immediately after the training, but its turnover decreased after 1 week of rest. In hypothalamus serotonin and 5-HIAA decreased immediately after training, followed by a rebound increase in their levels after 1 week of post-training rest.^[26]

Two studies^[23,24] used a spontaneous wheel running model (to avoid other stressors such as foot-shock) to examine changes in serotonin metabolism and turnover. An aromatic amino acid decarboxylase (AADC) inhibitor was used to measure 5-HTP (the direct precursor of serotonin) accumulation, which can give an estimate of serotonin synthesis.^[23] There were no statistically significant differences in 5-HTP. Chalouff et al.^[52] however, found regional differences in tryptophan

Table III. The effects of exercise on serotonin

Reference	Subjects	Exercise	Training	Brain area	Results
Barchas & Freedman ^[6]	S-D Male (200g) n = 50	Treadwheel: 3h, 1.8 m/min Swim to exhaustion: 15°C, 15-30 mins 23°C: 4 to 6h		Whole brain	↑ ↑ ↑ No difference serotonin or 5-HIAA
Speciale et al. ^[20]	S-D Male (250-300g) n = 6	Running wheel: 6.3 m/min, 1h		Striatum (n. caudatus)	5-HIAA ↑ [5-HIAA]/[serotonin] ↑ Serotonin unchanged in all regions
Heyes et al. ^[25]	S-D Male (350-400g) n = 25 (± 5/group)	Treadmill: 36 m/min RTE, or after 6, 11, 16.5 mins		Striatum Brainstem, hypothalamus	↓ Serotonin in all regions but only statistically sig. in striatum, cortex, midbrain
Lukaszyk et al. ^[38]	Wistar rats Male (150-220g) n = 5/group	Treadmill: 20 mins, 30 m/min		Cortex, hippocampus, hypothalamus, striatum, midbrain, cerebellum	↑ Serotonin, ~ 5-HIAA
Acworth et al. ^[42]	Male rats n = 23 (TTL 5/group) Wistar Rats Male (220g)	Treadmill: 90 mins, 25 m/min, 4° incline	1 week 4 to 5 times; last session 1h at 20 m/min	Whole brain	Serotonin ~ but ↑ serotonin metabolism 5-HIAA ↑ in 1h and 2h run
Chaoulouf et al. ^[49]	Wistar rats Male (250-300g) n = 5 (brain) n = 6 (CSF)	Treadmill: 60 mins, 20 m/min	1 week 4 to 5 times last session 1h at 20 m/min	Whole brain CSF withdrawal	↑ 5-HIAA indicating serotonin turnover, ↑ return to normal after 1h ↑ 5-HIAA
Chaoulouf et al. ^[51]	Wistar rats Male (± 200g) n = 4 groups of 10	Treadmill: 120 mins, 20 m/min	Short term: 1 week, 4 to 5 times Last session 1h at 20 m/min Long term: 8 weeks, 5 days/week, 1h/day, 20 m/min	Whole brain (minus cerebellum)	No effect from training on basal serotonin Serotonin = stable during exc (but sign. ↑ from long term) 5-HIAA ↑ (but sign. ↓ from long term) Serotonin ↓ during exc 5-HIAA ↑
Acworth et al. ^[42]	Male rats	Treadmill: 90 mins, 25 m/min, 4° incline	5 weeks, 6 days/week, 60 m/day, incline 1° to 4°	Whole brain	~ Serotonin ↑ 5-HIAA sign.
Chaoulouf et al. ^[43]	Wistar rats Male (220-250g) n = 5/group	Treadmill: 60 mins, 22 m/min	1 week 4-5 days/week, last session 1h, 20 m/min	Whole brain	↑ TRP ↑ Serotonin (NS) ↑ 5-HIAA
Bailey et al. ^[44]	Wistar rats Female n = 5	Treadmill: 60 mins, 20 m/min 5%	Treadmill accommodation: 3-4 weeks: speed + run time ↑, last session 20 m/min 5% 30 mins	Midbrain	TRP + exc: TRP ↑; 5-HIAA = ; serotonin = Fasting + exc: TRP ↑; 5-HIAA = ; serotonin = AMPH + exc: ↑ TRP; 5-HIAA ↓ (NS) AMPH + C: ↑ TRP; 5-HIAA ↑ (NS) α-MPT + exc: no effect on TRP and 5-HIAA in C; ratio 5-HIAA/TRP ↓ was prevented α-MPT; Haloperidol + C: no effect on serotonin Haloperidol: no effect on 5-HIAA in C & exc Exc + mCPP Serotonin ↑↑ 5-HIAA ↑↑

Table III. Contd

Reference	Subjects	Exercise	Training	Brain area	Results
Hoffmann et al. ^[24]	Spontaneously hypertensive rats	Running wheel Spontaneous running animals killed 1-2; 23-24 or 47-48h after last exc. period	5 week period 4 km/24h TTL: 7week	Limbic forebrain Brainstem Striatum, cortex, diencephalon	Serotonin & 5-HIAA ↓ sign. (1 to 2 h/run) - serotonin stayed sig. ↓ 5-HIAA ↓ sign. 48h post-run NS serotonin or 5-HIAA
Brown et al. ^[61]	Wistar rats Female n = 40	Treadmill ± 30 m/min	8 weeks, 5 days/week, 30 mins/day (normal diet or fat diet)	Whole brain Cerebellum Midbrain	Serotonin ↑ Serotonin ↑ Serotonin ↑
Hellhammer et al. ^[62]	Wistar rats n = 21	Running wheel	Spontaneous running	Pons-medulla Thalamus Hypothalamus Midbrain Septum Hippocampus Striatum	Serotonin ^a ↑, 5-HIAA ↓ No ↑ Serotonin ^a ↓, 5-HIAA ↓ Serotonin ^{a,b} ↓ Serotonin ^a ↑ Serotonin ^a ↓ (runners ↑ sed.) - 5-HIAA ↓ (runners-control) Serotonin ^{a,b} ↓ -Serotonin
Cicardo et al. ^[28]	Wistar rats Male n = 60	Swimming in 23°C: to exhaustion	Swimming in 23°C: to exhaustion	Whole brain	-Serotonin
Dey et al. ^[66]	Albino rats Male (200-250g) n = 50	Swimming: 60 mins, (35-36°C)	4 weeks, 6 days/week 30 min/day Sacrificed 24h or 7 days post exc	Cortex Hippocampus Hypothalamus Brainstem	Acute exc Serotonin = 5-HIAA ↑ NS Serotonin ↑ NS, 5-HIAA - Serotonin = 5-HIAA ↑ Serotonin ↑, 5-HIAA ↑ 24h training Serotonin ↑, 5-HIAA ↑ Serotonin ↓, 5-HIAA ↑ NS Serotonin = 5-HIAA ↓ Serotonin = 5-HIAA ↑
Wilson & Marsden ^[60]	Lister hooded rats Male (170-230g) n = 12 (exp + control)	Treadmill 60 mins 20m/min	4 weeks' accommodation speed & run time gradually increase	Cortex Hippocampus Hypothalamus Brainstem	7 Days post Serotonin ↑, 5-HIAA ↑ Serotonin = 5-HIAA ↓ Serotonin ↓, 5-HIAA ↓ Serotonin ↓, 5-HIAA ↓ Serotonin ↑ (NS) Serotonin ↑ (NS)
Bailey et al. ^[8]	Wistar rats Male (300-400g) TTL = 72 (n = 8/group)	Treadmill 60 mins or EXH 20 m/min	Treadmill: accommodation 3-4 weeks: speed + run time ↑, last session 20 m/mins, 5%, 30 mins	Midbrain Striatum Hypothalamus Hippocampus	1h Serotonin ↑, 5-HIAA ↑ Serotonin ↑, 5-HIAA ↑ Serotonin ↓, 5-HIAA ↑ Serotonin ↑ 5-HTP accum. ↑ =
Chaouloff et al. ^[52]	Wistar rats Male (220-250g) 6-7/group	Treadmill: 90 mins, 20 m/min	Treadmill accommodation: 1 week 4-5 day/week last session 1h, 20 m/min	Midbrain Striatum Hippocampus	Serotonin ↑ ↑ ↑ 5-HIAA ↑ ↑ ↑

Author	Subjects	Protocol	Whole brain			
			Serotonin	5-HIAA	Serotonin	5-HIAA
Romanowski & Gahric ^[53]	Wistar rats Male (220-250g) n = 10 + 10 control	Treadmill: 90 mins, 24 m/min	↑ 87% serotonin			
Blomstrand et al. ^[59]	Wistar rats Female (200-220g) n = ± 5/group	Treadmill to exhaustion: ± 17 mins; 7° incline, m/mins 30-35 m/min trained, ± 30-35 m/min	Sedentary: accommodation 10 mins/day to slow speed Training: 11 weeks, 6 days/week, 1 h/day, 30 m/min	Cortex Cerebellum Hippocampus Striatum Brainstem Hypothalamus	Serotonin Sedentary: No difference Trained: No difference	5-HIAA Sedentary: No difference Trained: No difference
Elam et al. ^[62]	Wistar Koyoto Male n = 25	Running wheel: spontaneous running animals were killed immediately or 24h after last running session	5 km/12h 7 days	Limbic forebrain, striatum, brainstem, cortex, spinal cord	No effect on 5-HTP accum.	No effect on 5-HTP accum.

a. In food restricted runners compared with control.
 b. Compared to not food restricted runners.
 Symbols and abbreviations: accum. = accumulation; αMPT = α-methyl-p-tyrosine; AMPH = amphetamine; C = control; exc = exercise; EXH = exhaustion; h = hour(s); inclin = inclination; mCPP = 5-HT_{1c} agonist; NS = not significant; RTE = run to exhaustion; serotonin = 5-hydroxytryptamine (5-HT); S-D = Sprague-Dawley rats; sig. = significant; TRP = tryptophan; TTL = total; 5-HIAA = 5-hydroxyindoleacetic acid; 5-HTP = 5-hydroxytryptophan; ↑ = increase; ↓ = decrease; - = no significant difference; = indicates no change.

utilisation into the serotonin synthesis pathway. Physical exercise differentially affected serotonin synthesis and metabolism in midbrain (cell bodies) and hippocampus and striatum (terminals). Running increased 5-HTP accumulation (after AADC inhibitor) in midbrain and decreased this accumulation in hippocampus while it was unaltered in striatum.

These results indicate that under some pharmacological conditions running causes region-specific alterations in the conversion of tryptophan into the serotonin synthesis pathway. If one considers that 5-HTP accumulation is a good index for serotonin synthesis, the authors hypothesise that this serotonin synthesis is increased in the midbrain and decreased in the hippocampus and unaltered in striatum of the exercising rat. Although the measurement of 5-HTP provides an estimate of the *in vivo* synthesis rates of brain monoamines, it does not enable an evaluation of the *in vivo* physiological activity of the monoamine pathways.^[23]

The other group that used spontaneous running animals, killed their animals 1 to 2 hours, 23 to 24 hours, or 47 to 48 hours, after the last training session.^[24] The serotonin and 5-HIAA levels in the limbic forebrain and brain stem decreased in the immediate post exercise period. A decrease in the tissue serotonin (as described by these authors) may thus indirectly suggest a relative increase in the release of serotonin.^[24] However, alterations in tissue levels of transmitters or their metabolites are crude measures of activity, which do not necessarily reflect corresponding changes in synaptic release. Recent microdialysis studies confirm the increased serotonin release during exercise.^[55,56,57]

Again it is difficult to draw conclusions from the studies that examined the influence of exercise on brain serotonin because there is no uniformity in study designs, exercise protocols, brain regions of interest and measuring methods. Even the strains of animals used are different.

We agree with Dunn and Dishmann^[13] who, in their review on exercise and the neurobiology of depression, point out some weak points of these studies.^[13,58] Briefly, most studies examined whole

brain neurotransmitter levels or have not assessed the same brain regions. The first studies used fluorescence spectrometry to determine transmitter levels, while later studies used more sensitive methods such as high performance liquid chromatography (HPLC) which allow measurement of multiple neurotransmitters and metabolites within the same sample.^[13] The discrepancies in measuring methods or statistical analysis prevents direct comparisons of results. For example, in the study of Blomstrand et al.^[39] dopamine level increased 65% in brainstem after exercise in trained rats. However, due to the large inter-individual variability of the responses, the authors could not reach statistical significance.^[39]

Most studies used forced locomotion on a treadmill,^[2,9,12,26,29,31,34,35,38,39,41-46,49,51-54,56] running wheel,^[16,17,19,20,25,30,33,59] spontaneous running,^[21,23,24,36,40] or swimming.^[16,18,21,28,32,37] Many studies used other stressors in addition to running or swimming. Swimming in water of different temperatures or the fear of drowning, or foot-shock during running could influence results.^[13] Animals that were tested after a single exercise session ran at different speeds. The chronic exercise protocols mostly used an accommodation period in which running speed and run time gradually increased, but these protocols varied from study to study. Only a few studies examined the effects of their training programme on endurance capacity or fitness level of the animals.

Measuring neurotransmitter levels in homogenates makes no distinction between extracellular and intracellular levels, and gives no indication of neurotransmitter release. Finally, measurement of single neurotransmitter levels does not provide much information on the relationships between neurotransmitters.

2.4 Neurotransmitter Interactions

Although it is difficult to compare the above mentioned studies, it seems that physical exercise influences the synthesis and metabolism of monoamines in various brain regions. There is evidence from recent microdialysis studies^[48,60-63] that there

is a reciprocal influence of various neurotransmitters in regulating their release. The various studies that examined the influence of exercise on brain neurotransmitters indicate that both central dopaminergic and serotonergic activity are influenced by exercise. Chaouloff et al.^[43] examined whether compounds known to affect dopamine activity in brain could modify the serotonin response in brain during exercise. The dopamine metabolism was increased in serotonin-rich regions.^[43] Administration of amphetamine, while increasing levels of tryptophan in brain, diminished the formation of 5-HIAA (ratio 5-HIAA to tryptophan). The relative inhibition of synthesis of serotonin induced by running, was thus potentiated by administration of amphetamine while α -methyl-*p*-tyrosine (inhibitor of catecholamine synthesis) prevented this effect of exercise, and haloperidol (dopamine antagonist) did not produce any significant change.

This possible interaction between brain serotonin and dopamine during exercise, was also proposed by Bailey et al.^[9,44,54] who examined the effects of increased serotonin activity on endurance performance and brain dopamine and serotonin turnover. They used several serotonin agonists and antagonists to examine run time to exhaustion and brain serotonin and dopamine levels. They found increased dopamine and DOPAC levels with running. At exhaustion, however, the dopamine and DOPAC levels were consistently lower than after 1 hour of exercise. Increased dopamine and DOPAC levels were significantly attenuated by *m*-chlorophenylpiperazine administration (*m*CPP is a 5-HT_{1C} agonist), indicating a possible impaired brain dopamine synthesis. When a general serotonin agonist (quipazine dimaleate) was administered, brain dopamine level significantly decreased at exhaustion in midbrain, and slightly decreased in striatum, hypothalamus and hippocampus. DOPAC significantly increased at exhaustion in hypothalamus and remained unchanged during exercise in the other brain regions. Administration of LY53857 (5-HT_{1C}/5-HT₂ antagonist) increased dopamine and DOPAC level during exercise in mid-

brain, striatum and hypothalamus, while it was unchanged in hippocampus.

It is possible that the interaction between brain serotonin and dopamine during prolonged exercise could play a regulative role in the onset of fatigue.^[9] However, it seems that this interaction could be region specific and that neuroendocrine factors also play an important role as several authors already demonstrated.^[9,13,64]

Future studies need to examine the region specific interactions between multiple neurotransmitters (including excitatory and inhibitory amino acid transmitters) during exercise, and the importance of various receptors, since single neurotransmitters can have both inhibitory and excitatory effects.^[13]

3. Neurotransmission and Exercise Performance

3.1 Tryptophan, Branched Chain Amino Acids and Neurotransmission

The variability of neurotransmitter release is regulated by a number of processes.^[65] One of these presynaptic processes which modulates neurotransmission is the change in neurotransmitter synthesis resulting from the metabolic consequences of eating or exercise.^[65] The biosynthesis of serotonin is tightly controlled by the activity of its rate-limiting enzyme tryptophan hydroxylase, so increases or decreases in its substrate, tryptophan, trigger increases or decreases in serotonin synthesis and metabolism.^[64] Tryptophan and the large neutral amino acids, including the branched chain amino acids (BCAAs) [valine, leucine, isoleucine] use the same carrier to enter the brain, and therefore are competitors for transport over the blood brain barrier. The blood level of free tryptophan or the ratio of free tryptophan to other large neutral amino acids is an important parameter for this competition.^[66-68]

Levels of circulating total and free tryptophan in plasma, and the ratio of free tryptophan to other large neutral amino acids, depends on several factors, e.g. the rate of lipolysis, the activity of hepatic

tryptophan pyrrolase, the uptake into the peripheral and central tissues.^[64] As free fatty acid levels increase during endurance exercise, the amount of tryptophan bound to albumin is reduced, increasing the level of free tryptophan in the blood. Other factors such as a high carbohydrate meal,^[67] insulin administration,^[68] administration of L-tryptophan,^[69] or a combination of these factors will increase the level of free tryptophan in plasma.^[70]

Since brain serotonin synthesis depends on the plasma level of tryptophan, treatments that elevate plasma tryptophan will promote accelerated serotonin synthesis and/or metabolism.^[15] Changes in neurotransmission caused by eating and by exercise can thus affect all of the behavioural and physiological functions that precursor-dependent neurons happen to subserve.^[65] Table IV summarises the studies that used precursor loading and/or pharmacological manipulation to influence exercise performance in human or animal models.

Serotonin has been shown to induce sleep, depress motor neuron excitability, influence autonomic and endocrine function and suppress appetite.^[10-12,71] This led several authors to propose the 'central fatigue hypothesis'.^[10-12] In order to examine this hypothesis Blomstrand et al.^[11,72,73] performed several studies. First, the changes in plasma levels of amino acids were examined during a marathon run and an army training programme.^[11] Both types of exercise caused a significant decrease in plasma level of BCAAs. The plasma level of free tryptophan was found to increase significantly during the race.

In two studies the effects of administration of BCAAs on mental and physical performance were examined.^[72,73] In one study mental performance in 6 female soccer players was studied.^[73] The study participants were given carbohydrate drinks with or without BCAAs. Plasma BCAAs were significantly decreased and plasma free tryptophan was significantly increased with the placebo carbohydrate drink. When ingesting the carbohydrate plus BCAA drink, amino acid levels were significantly increased and plasma free tryptophan was not significantly elevated. Mental performance

Table IV. Precursor loading and/or pharmacological manipulation, neurotransmission and exercise

Reference	Subjects	Manipulation	Exercise	Results
Jacobs & Eubanks ^[88]	S-D Male (± 400g) n = 64 + control	Serotonin inj. i.p. - s.c. 5-HTP inj. i.p. - s.c.	Crossing tilt-cage; activity measured during 3h	Activity ↓ in a dose dependent manner Serotonin more potent inhibitor than 5-HTP Behavioural changes following 5-HTP inj. may be attributable to peripheral effects of serotonin
Davis et al. ^[91]	Humans Male (n = 6)	Serotonin reuptake inhibitor (fluoxetine)	Cycling to exhaustion: 70% $\dot{V}O_{2max}$	Time to exhaustion ↓ with fluoxetine
Wilson et al. ^[90]	Humans Male (n = 7)	Serotonin reuptake inhibitor (paroxetine)	Cycling to exhaustion: 70% $\dot{V}O_{2max}$	Time to exhaustion sig. ↓ with paroxetine
Vergara et al. ^[79]	Wistar rats (± 380g) n = 34	Water, glucose or BCAA	Treadmill run to exhaustion: 16 m/min 5% incline	Time to exhaustion sig. ↓ in BCAA group compared to glucose-group
Bailey et al. ^[44]	Wistar rats Female (n = 8)	mCPP	Treadmill to exhaustion: 20 m/min 5% incline	RTE ↓ in a dose response manner by mCPP administration
Bailey et al. ^[54]	Wistar rats Male & female TTL = 36	QD LY 53857	Treadmill to exhaustion: 20 m/min 5% incline	QD: RTE ↓ in a dose response manner LY 53857: RTE only ↑ in the highest dose
Heyes et al. ^[34]	S-D Male (340-440g) n = 57	6-OHDA lesion Apomorphine Clonidine	Treadmill: 36 m/min, 6-OHDA lesioned rats	6-OHDA: decreased RTE Apomorphine: increased RTE Apomorphine in 6-OHDA rats: ↑ RTE Clonidine: no effect on RTE
Segura & Ventura ^[74]	Humans	L-TRP or placebo	Treadmill: (all out 100% $\dot{V}O_{2max}$)	↑ Running performance
Stensrud et al. ^[75]	Humans	L-TRP or placebo	Treadmill: (all out 100% $\dot{V}O_{2max}$)	No influence
Hillegaart et al. ^[94]	Rats	8-OHDPAT	Open field activity and treadmill running	No influence
Wilckens et al. ^[3]	Wistar rats Male	Different serotonin agonists and antagonists	Spontaneous running wheel activity after semi-starvation	Running wheel activity ↓ with 5-HT _{1c} agonists. This effect was abolished by antagonists with high 5-HT _{1c} affinity
Blomstrand et al. ^[73]	6 female soccer players	6% CHO + BCAA drinks	Soccer game	Free TRP/LNAAs ↓; Mental performance ↑ Free TRP/LNAAs ↓; Mental performance =
Blomstrand et al. ^[72]	Humans 25 male (TTL) 193 male (TTL) (blood samples in 26)	Placebo or BCAA drinks	30km cross-country marathon	Mental performance ↑ (BCAA) Mental performance = (placebo) Exc performance = BCAA - placebo (subdivision marathon times gave sig. difference)
Madsen et al. ^[79]	Humans 9 Male cyclists	CHO CHO + BCAA water	100km cycling trial	No influence on exc performance
MacLean et al. ^[80]	Humans 5 Male	Control or BCAA	Knee extension exc 60 mins	Suppression protein degradation in BCAA group
Van Hall et al. ^[81]	Humans 10 Male	6% CHO (placebo) 6% CHO BCAA 6% CHO L-TRP	Cycling to exhaustion at 70-75% max power output	No influence on exc performance
Lambert et al. ^[82]	Humans 8 Male	10% CHO 10% CHO + BCAA	4h cycling at 55% $\dot{V}O_{2max}$ + 40km time-trial	No influence on exc and mental performance
Galiano et al. ^[71]	Humans	6% CHO drink + BCAA	Cycling at 70% $\dot{V}O_{2max}$ up to 255 mins	No influence on exc, mental and endocrine performance
Bailey et al. ^[9]	Wistar rats Male (300-400g) TTL = 72 (n = 8/group)	Treadmill: 20 m/min	RTE: accommodation 3-4 weeks: speed + run time, last session, 20 m/min 5% 30 mins	QD (1 mg/kg): time to exhaustion ↓ LY 53857 (1.5 mg/kg): time to exhaustion ↑

De Meirlier ^[62,63]	Humans Male n = 10	DA agonist (D1/D2) Serotonin antagonist (5-HT _{1c} /5-HT ₂)	Maximal incremental exercise test	↑ Exc performance No influence on exc performance
Gerard ^[69]	S-D Male n = 10/group	(+) - AMPH different doses	Treadmill RTE different speeds	RTE ↑ with low doses RTE ↑ with high dose ↑ Endurance in fatigued animals (1.25 mg/kg dose)

Abbreviations: AMPH = amphetamine; BCAA = branched chain amino acids; CHO = carbohydrate drink; DA = dopamine; exc = exercise; h = hour(s); inj. = injection; i.p. = intraperitoneal; LNAA = large neutral amino acids; L-TRP = L-tryptophan; LY 53857 = serotonin antagonist; mCPP = 5-HT_{1c} agonist; 5-HTP = 5-hydroxytryptophan; QD = quipazine dimaleate serotonin agonist; RTE = run to exhaustion; s.c. = subcutaneous; S-D = Sprague-Dawley rats; sig. = significant; T.T. = total; ↑ = increase; ↓ = decrease; = indicates no change; 6-OHDA = 6-hydroxydopamine; 8-OHDPAT = 8-HT_{1a} agonist.

was significantly improved after the soccer match with the players that ingested the BCAAs, whereas mental performance was unchanged in all categories in the placebo carbohydrate drink group.^[73] The authors concluded that an intake of BCAAs in addition to a standard carbohydrate drink during exercise appears to affect mental alertness during and/or after exercise.^[73]

A similar experiment^[72] studied the effects of BCAA supplementation on endurance performance and psychological responses to a 30km and a marathon run. In the 30km trial, valine, leucine and isoleucine were significantly decreased in the placebo trial whereas BCAA supplementation resulted in a significant increase in these amino acids. In this study free tryptophan was not measured but the authors concluded that the increased availability of BCAAs would serve to maintain the free tryptophan/BCAA ratio at a lower level. Marathon run performance times were not significantly different when the placebo and BCAA group were compared. In order to get statistically significant differences the authors subdivided the group into faster (≤3.05 hours) and slower runners (between 3.05 and 3.30 hours). They did not provide a rationale as to why this cut-off time was chosen nor whether a similar trend would have been observed if other times had been selected.^[71]

Nutritional status was either not reported or controlled and only a subset of runners was assessed for plasma amino acid levels. Although BCAA supplementation improved mental performance after the soccer match and 30km run, the runners ingesting the placebo did not experience a decreased psychological status in comparison with pre-event mental performance. Consequently as performance was not evaluated during the soccer match or 30km run, there is no evidence to suggest that the increased mental performance after these events influenced performance capacity during the events.^[71]

The above studies used field experiments (soccer match, marathon run) that are difficult to control because of the variations in environmental and/or race conditions, which may affect physio-

logical responses to exercise and performance capacity.

Several other animal and human studies have examined this tryptophan- and serotonin-linked central fatigue hypothesis and the subsequent peripheral and central effects of endurance exercise.^[74-82] Davis et al.^[73] investigated the effects of carbohydrate feedings during prolonged exercise to fatigue on changes in plasma BCAAs, lactate, free fatty acids, insulin. These parameters could be important regulators of plasma tryptophan transport into the brain. The major finding of this study was that levels of plasma free tryptophan and free tryptophan/BCAAs and free fatty acids increased progressively during prolonged exercise until fatigue, while plasma BCAAs remained unchanged in the placebo group and decreased in the carbohydrate group. The changes in plasma free tryptophan, free tryptophan/BCAAs and free fatty acids were attenuated in a dose-dependent manner and fatigue was delayed when subjects consumed carbohydrate drinks.

Gailano et al.^[77] concluded that addition of small quantities of BCAAs to a typical sport drink may serve to maintain plasma BCAA levels throughout prolonged exercise, but does not appear to have any effect on psychological, endocrine, or performance responses during prolonged cycling. One study found that the supplementation of BCAAs suppressed the protein degradation during knee extension exercises for 60 minutes.^[80]

During exercise plasma levels of free tryptophan increase and it is proposed that this could be 'balanced' by raising the plasma level of BCAAs. Thus, according to the Newsholme hypothesis, supplementation with BCAAs will improve performance, while supplementation with tryptophan will have the opposite effect. This hypothesis was tested by several authors^[79,81,82] and the results indicate that oral supplementation with BCAAs or tryptophan significantly increases the plasma level of BCAAs. Neither time to exhaustion during cycling at 70 to 75% maximum oxygen uptake ($\dot{V}O_{2max}$),^[81] nor performance during 100km cycling in well-trained cyclists^[79] differed. So, nei-

ther a positive effect of BCAA supplementation, nor a negative effect of tryptophan supplementation on performance was found.

Van Hall et al.^[81] investigated whether ingestion of tryptophan or BCAAs could influence performance. They calculated the transport of tryptophan (free or bound) into the brain during prolonged exhaustive exercise and found that the effect of BCAAs and tryptophan ingestion seems to be independent of whether total or free tryptophan is considered available for transport.^[81] It was concluded that manipulation of tryptophan supply to the brain either has no additional effect upon serotonergic activity during prolonged exercise or that manipulation of serotonergic activity functionally does not contribute to mechanisms of fatigue.^[81]

Verger et al.^[78] examined the effects of administration of BCAAs versus glucose or water during acute exercise in the rat. They measured exercise time, blood insulin and glucose levels at exhaustion in the animals. The results showed that following ingestion of BCAA physical performance was lower and blood glucose levels between groups did not differ, while blood insulin level at exhaustion was higher with BCAAs than after glucose administration.^[78] It would have been interesting to have measured plasma free fatty acid level in this study and link this with the other parameters in order to get a more complete image of insulin, glucose, BCAAs and free fatty acid levels.

The decreases in BCAAs during prolonged exercise, as seen in the Blomstrand et al. studies,^[11,72,73] could also be the consequence of elevated plasma insulin levels. As previously mentioned, fluid intake was not totally controlled in these field studies, therefore it is likely that the study participants consumed carbohydrate-containing drinks. The effects of the above mentioned peripheral parameters and the link with insulin levels need further investigation since several studies have shown that not only insulin,^[65,83] but also amylin,^[84] facilitates brain tryptophan uptake and monoamine metabolism.^[85-87]

Taken together these studies indicate that fatigue can be delayed by ingesting carbohydrates,^[76] which are an important energy source for muscle and brain function,^[88] and that until today there is little evidence to support the hypothesis that BCAA supplementation will increase performance.

3.2 Precursor Loading, Pharmacological Manipulation and Neurotransmission

The possibility of a centrally mediated fatigue during exercise was discussed by Romanovski and Gabrielc^[53] in the mid-1970s. They linked serotonin to a possible inhibition of brain oxidoreductive processes, while others^[15,34,35,43] pointed out the role of dopamine in the onset of central fatigue. This brings us back to the possible interaction between neurotransmitters and their mutual influence on exercise performance. This was investigated by several authors who used precursor loading and/or pharmacological manipulation on exercise performance in animals and humans.

The effects of tryptophan supplementation on human performance were examined^[74] and it was hypothesised that administration of tryptophan before exercise could contribute to a decreased sense of discomfort and pain associated with prolonged exercise. During a treadmill test at 80% $\dot{V}O_{2max}$, the total exercise time to exhaustion increased by 49% in the tryptophan group, but this increase could have been confounded by a spectacular improvement in 2 of the 8 participants (160 and 260% increase in running time). This study was criticised by Chaouloff^[89] and Stensrud et al.,^[75] who performed a similar study and found no differences between tryptophan and placebo groups.

Two studies^[90,91] examined the effect of a serotonin reuptake inhibitor on time to exhaustion in humans. Both found a decrease in exercise time to exhaustion with the reuptake inhibitor compared to placebo. There were no differences in plasma glucose or lactate between both groups indicating the possible role of the central serotonergic systems in fatigue in people. De Meirleir^[92] examined the influence of a dopamine agonist (pergolide) and a

serotonin antagonist (ketanserin) on exercise performance. The results showed that oral treatment with the serotonin (5-HT_{1C}/5-HT₂) antagonist had no influence on exercise performance. It did not alter heart rate at rest or during exercise, but it elicited a shift to the right of the lactic acid curve.^[92] The dopamine agonist (D₁/D₂) lowered heart rate, systolic blood pressure and enhanced maximal work capacity.^[92,93]

Chaouloff et al.^[42,43,49-52] conducted a number of studies on the effects of tryptophan loading and/or exercise on central serotonin synthesis and metabolism. Under some pharmacological conditions running caused region-specific alterations in the conversion of tryptophan into the serotonin synthesis pathway.

The role of serotonin in the regulation of motor mechanisms is complex. Thus, a depletion of brain and spinal serotonin, as well as an increase in the availability of central serotonin, can result in a decrease or an increase in motor activity depending on the experimental model used.^[94]

There are numerous levels at which central serotonin can affect motor behaviour, from sensory perception, sensory-motor integration to motor effector mechanisms. For reviews of the involvement of serotonin in the initiation or modulation of motor patterns, neuromuscular function and motor control, see Jacobs and Fornal,^[6] Jacobs,^[7] and Wallis.^[95] Jacobs and Eubanks^[96] measured the effects of serotonin and 5-HTP injections on motor activity and found it to decrease in a dose-dependent manner, with serotonin being a more potent inhibitor than 5-HTP. They attributed these behavioural changes to peripheral effects of serotonin. These results were not confirmed by Wilckens et al.,^[3] who did not find any effect of serotonin administration on running wheel activity.

Another group^[94] investigated the effects of systemically administered 8-OH-DPAT (a serotonin agonist with preference for the 5-HT_{1A} binding site) on motor activity in open-field locomotion, and on treadmill running in rats. The spontaneous locomotor activity and rearing (vertical activity) were dose-dependently decreased by the adminis-

tration of the 5-HT_{1A} agonist. There were no statistically significant effects by 8-OH-DPAT on treadmill locomotion, i.e. the motor coordination was intact.^[94]

Chaouloff^[117] examined whether a short exercise training programme (4 days) could influence 5-HT_{1A} receptor-mediated behaviours. The effects of 8-OH-DPAT administration were not affected by training or acute exercise.^[117] It should be mentioned that the administration of 8-OH-DPAT and other drugs including precursors, agonists and releasers, produces various signs of the so-called 'serotonin syndrome'.^[7] This syndrome is characterised by hyperactivity, head shakes or 'wet dog' shakes, hyper-reactivity, tremor, rigidity, hind-limb abduction, Straub tail, lateral head weaving and reciprocal forepaw treading. These behavioural signs are sometimes used as an indication for central serotonin activity.^[7]

Wilckens et al.^[3] found with the same agonist (8-OH-DPAT) that locomotion increased during the first and second hour at the lowest dose and that at the higher doses locomotion was inhibited. This suppressed locomotion at higher doses could be a consequence of a behavioural impairment resulting from the 'serotonin syndrome'.^[3] They further evaluated the effects of serotonin receptor agonists and antagonists with selectivity for various serotonin receptor subtypes on running wheel activity in the rat model for semi-starvation-induced hyperactivity, where running wheel activity was stabilised for 10 weeks at a high level of circa 20 to 25 km/day. The results showed that excessive running in the semi-starved rat is suppressed by activation of the 5-HT_{1C} receptors and that activation of pre-synaptic 5-HT_{1B} receptors resulted in a decreased serotonin release.^[3]

These results were confirmed by Bailey et al.^[44] who found a decreased run time to exhaustion in a dose-response manner in animals treated with mCPP (5-HT_{1C} agonist). Other studies^[97,98] also suggested that hypolocomotion induced by mCPP could be mediated via postsynaptic 5-HT_{1C} receptors.

That 5-HT_{1C} receptors play a major role in the development of compulsive running is supported by the fact that the effect of the 5-HT_{1C} receptor agonists on wheel running in rats could only be counteracted by serotonin antagonists which have high affinity for the 5-HT_{1C} receptor (metergoline and mianserin).^[3] The inhibitory effect of the agonists on running wheel activity was prevented by pretreatment with antagonists that also had high affinity for the 5-HT_{1C} receptors.^[3] Bailey et al.^[9,54] examined the effects of quipazine dimaleate (QD), a general serotonin agonist with high affinity for the 5-HT₃ receptor^[3] and LY53857, a serotonin antagonist specific to 5-HT_{1C} and 5-HT₂ receptors. Run time to exhaustion was reduced in a dose-dependent manner by increasing dosages of quipazine dimaleate, while the time to exhaustion was increased with LY53857 administration but only at the highest dose.^[54] The results further indicated that QD appeared to block the increase in dopamine and DOPAC after 1 hour of exercise and LY53857 prevented the decrease in dopamine and DOPAC at fatigue,^[9] indicating the importance of the interaction between brain serotonin and dopamine in the onset of fatigue.

Pretreatment of exercising rats with amphetamine,^[99] a dopamine releaser, or apomorfine^[34] (a dopamine agonist), extends the time to exhaustion. Heyes et al.^[34] used the 6-hydroxydopamine (6-OHDA) model to induce a dopamine lesion and found these animals to have significant shorter run times to exhaustion. When apomorfine was given to 6-OHDA lesioned rats, their run time increased compared with saline-administered lesioned rats. Clonidine (noradrenaline receptor agonist) given to these animals had no effect.

These studies indicated that central dopamine depletion hastens time to exhaustion, while increasing central dopaminergic activity prolongs time to exhaustion.^[34] In a follow-up study the same authors^[35] found an increase in both dopamine synthesis and release with exercise. Striatal dopamine depletion had no effect on the rats ability to run during the first 40% of an exhaustion run, but accelerated the deterioration in exercise perfor-

mance during the remaining 60% of the run and hastened exhaustion.

The results of these studies^[34,35] suggest that the time to exhaustion is influenced by the activity of nigrostriatal dopaminergic neurons. During the later phase of the run to exhaustion there is probably a need to increase striatal dopaminergic activity. If this is the case, dopaminergic agonists may improve exercise capacity by facilitating such recruitment.^[35] On the other hand, these authors state that dopamine accumulation in the striatum late in exercise as found in their studies,^[34,35] may reflect decreases in dopamine release, perhaps due to activation of dopamine autoreceptors.^[35]

A low dose of haloperidol (dopamine antagonist) disrupts the treadmill performance^[100] and Chaouloff et al.^[43] reported that when haloperidol was administered at the beginning of exercise the animals were unable to run. The injection of haloperidol at the end of exercise caused a large increase in DOPAC in the brains of controls and runners.^[43]

The results from these studies emphasise the importance of dopamine and serotonin (and probably other neurotransmitter) interactions during exercise. Central 5-HT₃ receptors might be involved in the observed behaviour because they interact with the dopaminergic neurotransmitter system.^[3] LY53857 is a potent and selective 5-HT₂ receptor antagonist, but has also affinity for α_2 -receptors, therefore at the highest dose can interact with catecholaminergic receptors^[101] since Wilckens et al.^[3] found that propranolol increased motor activity in the rats, which could arise from the action of this serotonin antagonist (5-HT_{1A-B-C}) on β -adrenoceptors.

Taken together, these results indicate a role for several dopamine and serotonin transmitter receptors in motor control and the so-called 'central fatigue', but as Bailey and Davis^[102] pointed out: any possible role of serotonin, dopamine (and other transmitters) in motor function should be perceived as a continuum. This continuum is not only important at the brain level, but has its own importance in the interaction between central neurotrans-

mission and the peripheral processes during exercise, including the neuro-endocrine system, especially the activity of the hypothalamic-pituitary-adrenal (HPA) axis. Neurotransmitter systems not only influence each other, but they also are intimately linked to the HPA axis.^[13,58,64] We will not include an overview of the interactions between stress hormones and central neurotransmitter systems, for a review on serotonin and stress hormones see the excellent review of Chaouloff.^[64]

4. Measurement of Extracellular Neurotransmitter Levels

The review of the literature demonstrates that serotonergic, noradrenergic and dopaminergic neuronal systems are influenced in different ways during exercise. However, most of the studies were post mortem experiments which used indirect measurements such as the ratio of neurotransmitter to metabolites, or precursor to neurotransmitter to predict neurotransmitter release during exercise. Changes in the brain content of monoamine transmitters with tissue assay are now regarded as a rather inaccurate method to estimate changes in the release rate of these transmitters.^[103] Recently, new techniques such as microdialysis^[104-106] and voltammetry^[103,107] were introduced to measure *in vivo* release of neurotransmitters (see table V).

The voltammetry method is based on the application of a potential to an electrode in a conducting solution.^[103] The electrodes are implanted in the brain and an oxidation current is generated as molecules in the extracellular fluid are oxidised at the electrode surface.^[103]

Microdialysis is a means of assessing alterations in neurotransmitter release in brain extracellular space. It can collect virtually any substance from the brains of freely moving animals with a limited amount of tissue trauma.^[106] This method allows the measurement of local neurotransmitter release in combination with on-going behavioural changes such as exercise. A number of microdialysis studies have documented changes in extracellular neurotransmitters in several brain areas dur-

Table V. Effects of exercise on extracellular neurotransmitter levels

Reference	Subjects	Exercise	Training	Brain area	Methods	Results
Haltori et al. ^[113]	Wistar rats n = 34 TTL (4 groups)	Treadmill: 20 mins 18 m/min	7 days' accommodation 20 mins, 18 m/min	Striatum	Microdialysis	DA ↑, DOPAC ↑, HVA ↑
Sabol et al. ^[113]	Wistar rats Male (300-390g) n = 12	Circular disk, treadmill: 24 mins 10 m/min, water reinforcement		Lateral striatum Medial striatum + n. accumbens Parietal lobe of cerebral cortex	Microdialysis	DOPAC ↑ no sig, ↑ DA DOPAC ↑ no sig, ↑ DA ACh ↑ NA ↑, serotonin ↑
Kurosawa et al. ^[57]	Wistar rats Male (300-390g) n = 12	Treadmill: 2.3 m/min, manually moved			Microdialysis	
Pagliari et al. ^[111]	Rats	Treadmill: 60 mins, 25 m/min, 3% incl.	2 weeks, 5 days/week, 1 h/day 25 m/min 3% incl.	Frontal cortex	Microdialysis	NA ↑ MHPG-DHPG ↑ Serotonin ↑
Wilson & Marsden ^[56]	Lister-hooded rats Male (170-230g) n = 5	Treadmill: 60 mins, 20 m/min	4 weeks accommodation: speed & run time gradually increase	Hippocampus	Microdialysis	
Meeusen et al. ^[55-114]	Wistar rats Male (250-300g) n = 10	Treadmill: 20 mins, 12 m/min	2 weeks accommodation; 2 days/week: speed & run time gradually increase	Striatum	Microdialysis	DA ↑ NA ↑ Serotonin ↑ GABA ~ GLU ↑
Geim et al. ^[115]	S-D Male n = 6	Treadmill: 60 mins, 26.6 m/min	Accommodation 5 days/week	Ventral horn, spinal cord	Microdialysis (chronic implantation)	No ↑ serotonin during exc.
Berfollucci-D'Angio et al. ^[23]	S-D Male (250-300g) n = 5	Rotarod: 40 mins forced locomotion 5 m/min	5 days' accommodation	Striatum n. accumbens Prefrontal cortex	Voltammetry	DOPAC ↑ DOPAC ↑
Guadalupe et al. ^[116]	Male rats n = 10	Running wheel: motor driven	10 mins 3m to 5.4 m/m/n	n. accumbens: shell core	Voltammetry	DA~ DOPAC~

Abbreviations and symbols: ACh = acetylcholine; DA = dopamine; DHPG = 3,4-dihydroxyphenylglycol; DOPAC = 3,4-dihydroxyphenylacetic acid; exc = exercise; GABA = γ-aminobutyric acid; GLU = glutamate; h = hour(s); HVA = homovanillic acid; MHPG = 3-methoxy-4-hydroxyphenylethylglycol; NA = noradrenaline (norepinephrine); S-D = Sprague-Dawley rats; sig. = significant; TTL = total; ↑ = increase; ↓ = decrease; ~ = no significant difference.

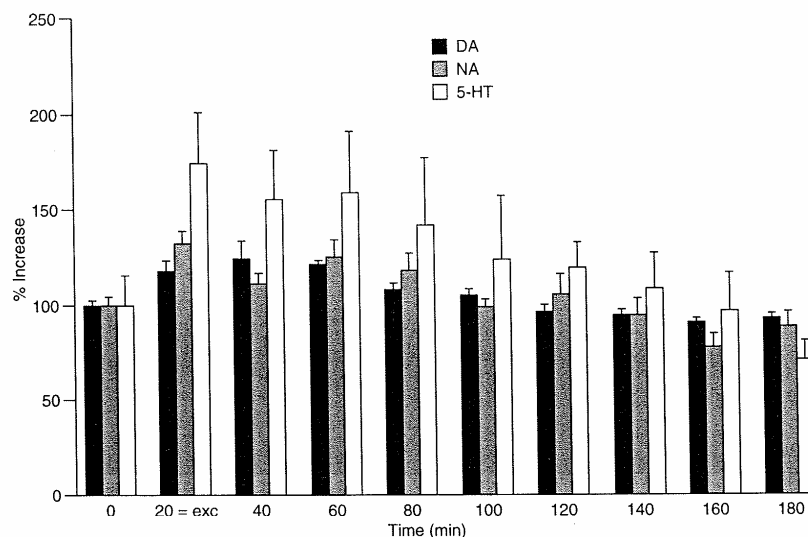


Fig. 3. The effects of 20 minutes of exercise on extracellular monoamine levels in rat striatum (after Meeusen et al. 1994^[55], with permission). Compared to baseline values, the brain monoamine release increased during 20 minutes of exercise. Maximal extracellular levels were obtained 20 minutes post-run for dopamine, while noradrenaline (norepinephrine) and serotonin (5-hydroxytryptamine; 5-HT) values peaked during running. After this increase extracellular levels of the neurotransmitters remained above baseline for at least 60 to 80 minutes, while serotonin remained elevated up to 120 minutes after the exercise was stopped. *Abbreviations:* DA = dopamine; NA = noradrenaline (norepinephrine).

ing exposure of the animals to different stimuli.^[47,108,109]

Recently, the first reports appeared, demonstrating that it is possible to measure extracellular levels of neurotransmitters with microdialysis in the rat brain during exercise and recovery from exercise.^[55,56,110,111,114] Two studies^[55,112] found that 20 minutes of exercise on a treadmill, significantly increased dopamine release in rat striatum. Hattori et al.^[112] combined microdialysis with running in order to evaluate motor deficit and improvement following dopaminergic grafts in 6-OHDA lesioned rats. Dopamine, DOPAC and homovanillic acid significantly increased during the treadmill exercise in their control animals.^[112]

In another study a circular treadmill (speed of the treadmill circa 10 m/min) was used in order to let the animals turn and 'walk in place' for sucrose water reward.^[113] The rats were fixed by their tails and walked in place for 24 minutes. Extracellular

levels of dopamine and DOPAC were measured in the nucleus accumbens/medial striatum and lateral striatum. An increase in dopamine, and DOPAC release in the lateral striatum was found. The authors were unable to conclude whether these changes were due to motor activity, the act of drinking, the tail pinch stress or the amount of fluid consumed.

Another study registered the simultaneous release of monoamines in rat striatum, during and following exercise, and found a significant increase of these neurotransmitters (fig. 3).^[55] In a similar experiment^[114] it was demonstrated that a light exercise regimen is able to significantly increase extracellular levels of glutamate (GLU), while γ -aminobutyric acid (GABA) remains unchanged (fig. 4). These results could indicate the existence of a functional interaction of several brain neurotransmitters in the regulation of locomotion.

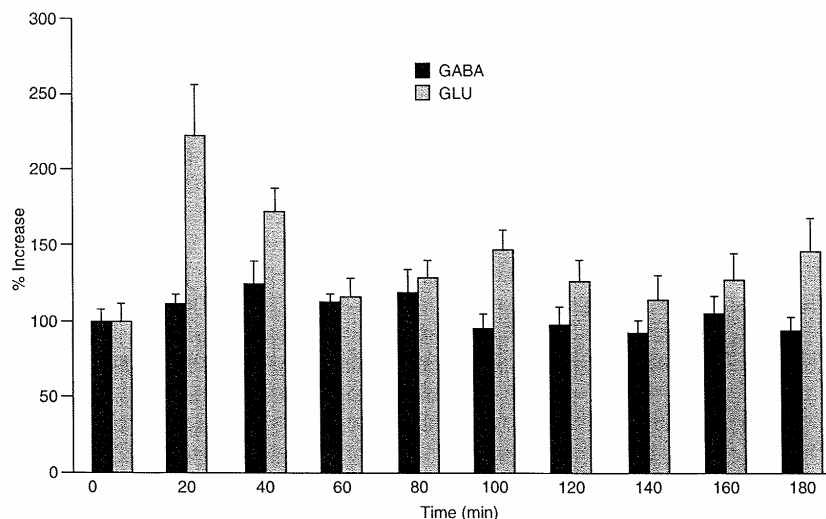


Fig. 4. The effects of 20 minutes of exercise on extracellular γ -aminobutyric acid (GABA) and glutamate (GLU) levels in rat striatum.^[114] Compared to baseline, GLU levels increased significantly during exercise, and remained significantly elevated during the following 20 minutes. GABA levels showed no significant increase.

Kurosawa et al.^[57] used a treadmill that was manipulated manually at a low speed (2.3 m/min). The animals were restrained in a metal harness which was fixed to the rat's chest and abdomen with plaster. Extracellular acetylcholine (ACh), noradrenaline and serotonin in the parietal lobe of the cerebral cortex was examined. Walking for 5 minutes produced an increase of all 3 neurotransmitters.

Treadmill running for 60 minutes significantly increased extracellular serotonin level in the hippocampus of trained rats.^[56] Pagliari et al.^[111] examined the effect of exercise on the *in vivo* cerebral release and turnover of noradrenaline in trained rats running on a treadmill for 60 minutes. The authors used a chronic probe implantation in the frontal cortex. Noradrenaline turnover and release increased during exercise and even further increased when exercise time was prolonged to 2 hours of running.

Gerin et al.^[115] used an interesting approach. To investigate the effects of exercise on spinal cord serotonin, these authors chronically implanted a microdialysis probe in the ventral horn of the lum-

bar spinal cord of rats. The probe was kept in place during 40 days. Extracellular release of serotonin did not increase during 60 minutes of exercise.

Two studies^[22,116] used *in vivo* voltammetry. Bertolucci-D'Angio et al.^[22] studied the dopamine metabolism in different brain regions and compared forced locomotion with several other stressors. Forced locomotion of rats on a rotarod for 40 minutes increased the amplitude of the DOPAC oxidation peak in the striatum and the nucleus accumbens, but failed to affect the DOPAC peak in the prefrontal cortex. This increase in the striatum and nucleus accumbens is compatible with the currently held view that the nigrostriatal dopaminergic neurons are associated with motor function.^[22]

In addition to these results Guadeloupe et al.^[116] investigated the effects of forced locomotion and spontaneous locomotion on dopamine and DOPAC in the nucleus accumbens. They found that continuous and intermittent locomotion increased the levels of dopamine and its metabolite indicating the involvement of the nucleus accumbens in the

initiation but not the maintenance of movement.^[116]

The recently developed and more sophisticated *in vivo* methods such as microdialysis and *in vivo* voltammetry will improve our insight into the relationship between the monoamine and other transmitters during on-going behaviour such as exercise. These methods will allow us to monitor extracellular release and metabolism of various neurotransmitters. However, the few studies that have been published up till now, already show the same discrepancies as in the post mortem studies. They used different exercise models, for example, with or without extra stress (electrical grid at the end of the treadmill, restraint in a harness), other running speeds and training regimens. We therefore hope that in the future these precise collection methods, will be used in well-defined experimental protocols in order to being able to compare the different results.

5. Conclusions

There is consensus that monoaminergic neurons are involved in a number of functions that regulate locomotion. Although most studies used different experimental protocols, it can be concluded that brain neurotransmission is influenced by exercise. The effects of exercise on neurotransmission should be explored in a multidimensional way because there is a constant interaction between several neurotransmitters and their respective receptors during locomotion.

Many neurotransmitters or neuromodulators influence an individual's ability to exercise via actions in both the peripheral and central nervous system. The intracerebral mechanisms responsible for the central fatigue phenomenon have not been fully identified. Animal and human studies will help us to find out the effects of various pharmacological manipulations on central fatigue.

It would be of interest to study whether the different neurotransmitter interactions can influence 'central fatigue' during exercise. We need to develop standard experimental strategies in order to examine the effects of exercise on brain functions.

Neurotransmitter inter-relationships are important since these interactions reflect the multidimensional image of the different processes that happen in the brain during exercise. *In vivo* methods will allow us to explore the numerous interactions between neurotransmitters and receptors and get more insight into areas such as neurotransmitter release, metabolism, reuptake and receptor sensitivity.

Acknowledgements

The authors gratefully acknowledge Ilse Smolders for her valuable remarks during preparation of this manuscript. We further acknowledge the skilful technical assistance of Angeline De Troyer. The personal studies cited in this work were partially supported by a grant from the research council of the Vrije Universiteit Brussel.

References

1. Fentem P. Benefits of exercise in health and disease. *BMJ* 1994; 308: 1291-5
2. Freed C, Yamamoto B. Regional brain dopamine metabolism: a marker for the speed, direction and posture of moving animals. *Science* 1985; 229: 62-5
3. Wilckens T, Schweiger U, Pirke K. Activation of 5-HT_{1C}-receptors suppresses excessive wheel running induced by semi-starvation in the rat. *Psychopharmacol* 1992; 109: 77-84
4. Gil M, Marti J, Armario A. Inhibition of catecholamine synthesis depresses behaviour of rats in the holeboard and forced swim tests: influence of previous chronic stress. *Pharmacol Biochem Behav* 1992; 43: 597-601
5. Hassler R. Striatal control of locomotion, intentional actions and of integrating and perceptive activity. *J Neurol Sci* 1978; 36: 187-224
6. Jacobs B, Fornal C. 5-HT and motor control: a hypothesis. *Trends Neurosci* 1993; 16 (9): 346-50
7. Jacobs B. Serotonin and behaviour: emphasis on motor control. *J Clin Psychol* 1991; 52: 12 Suppl.: 17-23
8. Marsden C. The mysterious motor function of the basal ganglia. The Robert Wartenberg Lecture. *Neurology* 1982; 32: 514-39
9. Bailey S, Davis J, Ahlborn E. Neuroendocrine and substrate responses to altered brain 5-HT activity during prolonged exercise to fatigue. *J Appl Physiol* 1993; 74 (6): 3006-12
10. Newsholme E, Acworth I, Blomstrand E. Amino acids, brain neurotransmitters and a functional link between muscle and brain that is important in sustained exercise. In: Benzi G, editor. *Advances in myochemistry*. London: John Libby Eurotext, 1987: 127-38
11. Blomstrand E, Celsing F, Newsholme E. Changes in plasma levels of aromatic and branched chain amino acids during sustained exercise in man and their possible role in fatigue. *Acta Physiol Scand* 1988; 133: 115-21
12. Acworth I, Nicolass J, Morgan B, et al. Effect of sustained exercise on concentrations of plasma aromatic and branched chain amino acids and brain amines. *Biochem Biophys Res Commun* 1986; 137 (1): 149-53
13. Dunn A, Dishmann R. Exercise and the neurobiology of depression. *Exerc Sport Sci Rev* 1991; 19: 41-98

14. Herregodts P. Neurochemical studies of monoaminergic neurotransmitters in the central nervous system. Brussels: VUB Press, 1991
15. Chaouloff F. Physical exercise and brain monoamines: a review. *Acta Physiol Scand* 1989; 137: 1-13
16. Barchas J, Freedman D. Brain amines: response to physiological stress. *Biochem Pharmacol* 1963; 12: 1232-35
17. Gordon R, Spector A, Sjoerdsma A, et al. Increased synthesis of norepinephrine and epinephrine in the intact rat during exercise and exposure to cold. *J Pharmacol Exp Ther* 1966; 153: 440-7
18. Moore K, Larivière E. Effects of stress and d-amphetamine on rat brain catecholamines. *Biochem Pharmacol* 1964; 13: 1098-100
19. Moore K. Development of tolerance to the behavioural depressant effect of alpha-methyltyrosine. *J Pharm Pharmacol* 1968; 20: 805-6
20. Speciale S, Miller J, McMillen B, et al. Activation of specific central dopamine pathways: locomotion and footshock. *Brain Res Bull* 1986; 16: 33-8
21. Bliss E, Aillion J. Relationship of stress and activity on brain dopamine and homovanillic acid. *Life Sci* 1971; 10: 1161-9
22. Bertolucci-D'Angio M, Serrano A, Scatton B. Differential effects of forced locomotion, tail pinch, immobilisation and methyl-beta-carboline carboxylate on extracellular DOPAC levels in the rat striatum, nucleus accumbens, and prefrontal cortex: an in vivo voltammetric study. *J Neurochem* 1990; 55: 1208-15
23. Elam M, Svensson T, Thoren P. Brain monoamine metabolism is altered in rats following spontaneous long-distance running. *Acta Physiol Scand* 1987; 130: 313-6
24. Hoffmann P, Elam M, Thoren P, et al. Effects of long lasting voluntary running on the cerebral levels of dopamine, serotonin and their metabolites in the spontaneously hypertensive rat. *Life Sci* 1994; 54 (13): 855-61
25. De Castro J, Duncan G. Operantly conditioned running: effects on brain catecholamine concentrations and receptor densities in the rat. *Pharmacol Biochem Behav* 1985; 23: 495-500
26. Dey S, Singh R, Dey P. Exercise training: significance of regional alterations in serotonin metabolism of rat brain in relation to antidepressant effect of exercise. *Physiol Behav* 1992; 52: 1095-9
27. Dey S. Physical exercise as a novel antidepressant agent: possible role of serotonin receptor subtypes. *Physiol Behav* 1994; 55 (2): 323-9
28. Cicardo V, Carbone S, De Rondina D, et al. Stress by forced swimming in the rat, effects of misanserin and moclobemide on GABAergic and monoaminergic systems in the brain. *Comp Biochem Physiol C* 1986; 83 (1): 133-5
29. Sheldon M, Sorcher S, Smith C. A comparison of the effects of morphine and forced running upon the incorporation of ¹⁴C-tyrosine into ¹⁴C-catecholamines in mouse brain, heart and spleen. *J Pharmacol Exp Ther* 1975; 193: 564-75
30. Brown B, Van Huss W. Exercise and rat brain catecholamines. *J Appl Physiol* 1973; 34 (5): 664-9
31. Brown B, Payne T, Kim C, et al. Chronic response of rat brain norepinephrine and serotonin levels to endurance training. *J Appl Physiol* 1979; 46 (1): 19-23
32. Östman I, Nybäck H. Adaptive changes in central and peripheral noradrenergic neurons in rats following chronic exercise. *Neurosci* 1976; 1: 41-7
33. Stone E. Accumulation and metabolism of norepinephrine in rat hypothalamus after exhaustive stress. *J Neurochem* 1973; 21: 589-601
34. Heyes M, Garnett E, Coates G. Central dopaminergic activity influences rats ability to run. *Life Sci* 1985; 36: 671-7
35. Heyes M, Garnett E, Coates G. Nigrostriatal dopaminergic activity is increased during exhaustive exercise stress in rats. *Life Sci* 1988; 42: 1537-42
36. Rea M, Hellhammer D. Activity wheel stress changes in brain norepinephrine turnover and the occurrence of gastric lesions. *Psychother Psychosom* 1984; 42: 218-23
37. Sudo A. Time course of the changes of catecholamine levels in rat brain during swimming stress. *Brain Res* 1983; 276: 372-4
38. Lukaszuk A, Buckzo W, Wisniewski K. The effect of strenuous exercise on the reactivity of the central dopaminergic system in the rat. *Pol J Pharmacol Pharm* 1983; 35: 29-36
39. Blomstrand E, Perret D, Parry-Billings M, et al. Effect of sustained exercise on plasma amino acid concentrations and on serotonin metabolism in six different brain regions in the rat. *Acta Physiol Scand* 1989; 136: 473-81
40. Broocks A, Liu J, Pirki K. Semi-starvation induced hyperactivity compensates for decreased norepinephrine and dopamine turnover in the mediobasal hypothalamus of the rat. *J Neural Transm* 1990; 79: 113-24
41. Brown B, Piper E, Riggs C, et al. Acute and chronic effects of aerobic and anaerobic training upon brain neurotransmitters and cytochrome oxidase activity in muscle [abstract]. *Intern J Sports Med* 1992; 13: 92-3
42. Chaouloff F, Laude D, Guezennec Y, et al. Motor activity increases tryptophan, 5-hydroxyindoleacetic acid, and homovanillic acid in ventricular cerebrospinal fluid of the conscious rat. *J Neurochem* 1986; 46: 1313-6
43. Chaouloff F, Laude D, Meringo D, et al. Amphetamine and alpha-methyl-p-tyrosine affect the exercise induced imbalance between the availability of tryptophan and synthesis of serotonin in the brain of the rat. *Neuropharmacol* 1987; 26 (8): 1099-106
44. Bailey S, Davis J, Ahlborn E. Effect of increased brain serotonergic activity on endurance performance in the rat. *Acta Physiol Scand* 1992; 145: 75-6
45. Gilliam P, Spirduso W, Martin T, et al. The effects of exercise training on (³H)-spiperone binding in rat striatum. *Pharmacol Biochem Behav* 1984; 20: 863-7
46. MacRea P, Spirduso W, Cartee G, et al. Endurance training effects on striatal D₂ dopamine-receptor binding and striatal dopamine metabolite levels [letter]. *Neurosci* 1987; 79: 138-44
47. Boldry R, Willins D, Wallace L, et al. The role of endogenous dopamine in the hypermobility response to intra-accumbens AMPA. *Brain Res* 1991; 559: 100-8
48. O'Connor W, Morari M, Fuxe K, et al. Dopamine and NMDA receptor regulation of striatal GABA output neurons. In: Louilot A, Durkin T, Spampinato U, et al. editors. Monitoring molecules in neuroscience. Gradignan: Publi Typ 1994; 289-90
49. Chaouloff F, Elghozi J, Guezennec Y, et al. Effects of conditioned running on plasma, liver and brain tryptophan and on brain 5-hydroxytryptamine metabolism in the rat. *Br J Pharmacol* 1985; 86: 33-41
50. Chaouloff F, Kennett G, Serrurier B, et al. Amino acid analysis demonstrates that increased plasma free tryptophan causes the increase of brain tryptophan during exercise in the rat. *J Neurochem* 1986; 46: 1647-50
51. Chaouloff F, Laude D, Elghozi J. Brain serotonin response to exercise in the rat: the influence of training duration. *Biog Amines* 1987; 4: 99-106

52. Chaouloff F, Laude D, Elghozi J. Physical exercise: evidence for differential consequences of tryptophan on 5-HT synthesis and metabolism in central serotonergic cell bodies and terminals. *J Neural Transm [Gen Sect]* 1989; 78: 121-30
53. Romanowski W, Grabiec S. The role of serotonin in the mechanism of central fatigue. *Acta Physiol Pol* 1974; 25: 127-34
54. Bailey S, Davis J, Ahlborn E. Serotonergic agonists and antagonists affect endurance performance in the rat. *Intern J Sports Med* 1993; 14 (6): 330-3
55. Meeusen R, Sarre S, Michotte Y, et al. The effects of exercise on neurotransmission in rat striatum, a microdialysis study. In: Louilot A, Durkin T, Spampinato U, et al., editors. *Monitoring molecules in neuroscience*. Gradignan: Publi Typ 1994; 181-2
56. Wilson W, Marsden C. The effect of running on brain serotonin. In: Louilot A, Durkin T, Spampinato U, et al., editors. *Monitoring molecules in neuroscience*. Gradignan: Publi Typ 1994; 223-4
57. Kurosawa M, Okada K, Sato A, et al. Extracellular release of acetylcholine, noradrenaline and serotonin increases in the cerebral cortex during walking in conscious rats. *Neurosci Lett* 1993; 161: 73-6
58. Dishman R. Biological psychology, exercise, and stress. *Quest* 1994; 46: 28-59
59. Hellhammer D, Hingten J, Wade S, et al. Serotonergic changes in specific areas of rat brain associated with activity - stress gastric lesions. *Psychosom Med* 1983; 45: 115-22
60. Imperato A, Angelucci L, Casolini P, et al. Repeated stressful experiences differently affect limbic dopamine release during and following stress. *Brain Res* 1992; 577: 194-9
61. Ferré S, Cortes R, Artigas F. Dopaminergic regulation of the serotonergic raphe-striatal pathway: microdialysis studies in freely moving rats. *J Neurosci* 1994; 14 (8): 4839-46
62. Ohta K, Fukuuchi Y, Shimazu K, et al. Presynaptic glutamate receptors facilitate release of norepinephrine and 5-HT as well as dopamine in the normal and ischemic striatum. *J Auton Nerv Sys* 1994; 49: S195-S202
63. Zocchi A, Pert A. Alterations in striatal acetylcholine overflow by cocaine, morphine, and MK-801: relationship to locomotor output. *Psychopharmacol* 1994; 115: 297-304
64. Chaouloff F. Physiopharmacological interactions between stress hormones and central serotonergic systems. *Brain Res Rev* 1993; 18: 1-32
65. Wurtman R, Lewis M. Exercise, plasma composition and neurotransmission. In: Brouns F, editor. *Advances in nutrition and top sport*. Med Sport Sci. Basel: Karger, 1991; 32: 94-109
66. Fernstrom J. Role of precursor availability in control of monoamine biosynthesis in brain. *Physiol Rev* 1983; 63 (2): 484-546
67. Fernstrom J, Wurtman R. Brain serotonin content: physiological dependence on plasma tryptophan levels. *Science* 1971; 173: 149-52
68. Fernstrom J, Wurtman R. Brain serotonin content: physiological regulation by plasma neutral amino acids. *Science* 1972; 178: 414-6
69. Sharp T, Bramwell S, Grahame-Smith D. Effect of acute administration of L-tryptophan in the release of 5-HT in rat hippocampus in relation to serotonergic neuronal activity: an in vivo microdialysis study. *Life Sci* 1992; 50: 1215-23
70. Hernandez L, Parada M, Baptista T, et al. Hypothalamic serotonin in treatments for feeding disorders and depression as studied by brain microdialysis. *J Clin Psych* 1991; 52 (12 Suppl.): 32-40
71. Kreider RB, Miriel V, Bertun E. Amino acid supplementation and exercise performance. *Sports Med* 1993; 16 (3): 190-209
72. Blomstrand E, Hassmen P, Ekblom B, et al. Administration of branched chain amino acids during sustained exercise - effects on performance and on plasma concentration of some amino acids. *Eur J Appl Physiol* 1991; 63: 83-8
73. Blomstrand E, Hassmen P, Newsholme E. Effect of branched chain amino acid supplementation on mental performance. *Acta Physiol Scand* 1991; 143: 225-6
74. Segura R, Ventura J. Effect of L-Tryptophan supplementation on exercise performance. *Int J Sports Med* 1988; 9: 301-5
75. Stensrud T, Holm H, Stromme S. L-Tryptophan supplementation does not improve running performance. *Int J Sports Med* 1992; 13 (6): 481-5
76. Davis M, Bailey S, Woods J, et al. Effects of carbohydrate feedings on plasma free tryptophan and branched chain amino acids during prolonged cycling. *Eur J Appl Physiol* 1992; 65: 513-9
77. Galiano F, Davis J, Bailey M, et al. Physiological, endocrine and performance effects of adding branched chain amino acids to a 6% carbohydrate electrolyte beverage during prolonged cycling [abstract]. *Med Sci Sports Exerc* 1991; 23: S14
78. Verger P, Aymard P, Cynobert L, et al. Effects of administration of branched chain amino acids versus glucose during acute exercise in the rat. *Physiol Behav* 1994; 55 (3): 523-6
79. Madsen K, Christensen D. Administration of glucose, glucose plus branched chain amino acids or placebo during sustained exercise and their effects on a 100 km bike performance [abstract]. Ninth International Conference Biochemistry of Exercise: 1994 July 21-26; Aberdeen, Scotland, 35
80. MacLean D, Graham T, Saltin B. Branched chain amino acid supplementation attenuates net protein degradation during exercise [abstract]. Ninth International Conference Biochemistry of Exercise: 1994 July 21-26; Aberdeen, Scotland, 51
81. Van Hall G, Raaymakers J, Saris W, et al. Ingestion of branched-chain amino acids and tryptophan during sustained exercise - failure to affect performance. *J Physiol*. In press.
82. Lambert M, Velloza P, Wilson G, et al. The effect of carbohydrate and branched chain amino acid supplementation on cycling performance and mental fatigue [abstract]. Ninth International Conference Biochemistry of Exercise: 1994 July 21-26; Aberdeen, Scotland; 53
83. Martin-Du Pan R, Mauron C, Glaeser B, et al. Effect of various oral glucose doses on plasma neutral amino acid levels. *Metabolism* 1982; 31 (9): 937-43
84. Chance W, Balasubramaniam A, Thomas I, et al. Amylin increases transport of tyrosine and tryptophan into the brain. *Brain Res* 1992; 593: 20-4
85. Kwok R, Juorio A. Facilitating effect of insulin on brain 5-hydroxytryptamine metabolism. *Neuroendocrinol* 1987; 45: 267-73
86. Shimizu H, Bray G. Effects of insulin on hypothalamic monoamine metabolism. *Brain Res* 1990; 510: 251-8
87. MacKenzie R, Trulsson M. Does insulin act directly on the brain to increase tryptophan levels? *J Neurochem* 1978; 30: 1205-8
88. Coggan A, Coyle E. Carbohydrate ingestion during prolonged exercise: effects on metabolism and performance. *Exerc Sport Sci Rev* 1991; 19: 1-40
89. Chaouloff F. About the effects of L-tryptophan on exercise performance: lacunae and pitfalls [letter]. *Int J Sports Med* 1989; 10: 383
90. Wilson W, Maughan R. Evidence for a possible role of 5-hydroxytryptamine in the genesis of fatigue in man: admin-

- istration of paroxetine, a 5-HT reuptake inhibitor, reduces the capacity to perform prolonged exercise. *Exper Physiol* 1992; (77): 921-4
91. Davis M, Bailey S, Jackson D. et al. Effects of a serotonin agonist during prolonged exercise to fatigue in humans [abstract]. *Med Sci Sports Exerc* 1993; 25 (5): S78
 92. De Meirleir K. Studies on cardiovascular drugs and (neuro)humoral substances in dynamic exercise [PhD thesis]. Brussels: Vrije Universiteit Brussel, 1985
 93. De Meirleir K, Gerlo F, Hollmann W, et al. Cardiovascular effects of pergolide mesylate during dynamic exercise. *Proceedings of the British Pharmacology Society* 1986; 633P
 94. Hillegaart V, Wadenberg M, Ahlenius S. Effects of 8-OH-DPAT on motor activity in the rat. *Pharmacol Biochem Behav* 1989; 32: 797-800
 95. Wallis D. 5-HT receptors involved in initiation or modulation of motor patterns: opportunities for drug development. *Trends Neurosci* 1994; 15: 288-92
 96. Jacobs B, Eubanks E. A comparison of the locomotor effects of 5-hydroxytryptamine and 5-hydroxytryptophan administered via two systemic routes. *Pharmacol Biochem Behav* 1974; (2): 137-9
 97. Kennett G, Curzon G. Evidence that mCPP may have behavioural effects mediated by central 5-HT_{1C} receptors. *Br J Pharmacol* 1988; 94: 137-47
 98. Lucki I, Ward H, Frazer R. Effect of 1-(m-chlorophenyl) piperazine and 1-(trifluoromethylphenyl) piperazine on locomotor activity. *J Pharmacol Exp Ther* 1989; 249: 155-64
 99. Gerald M. Effect of (+)-amphetamine on the treadmill endurance performance of rats. *Neuropharmacol* 1978; 17: 703-4
 100. Ahlenius S, Hillegaart V. Involvement of extrapyramidal motor mechanisms in the suppression of locomotor activity by antipsychotic drugs: a comparison between the effects produced by pre- and post-synaptic inhibition of dopaminergic neurotransmission. *Pharmacol Biochem Behav* 1986; 24: 1409-15
 101. Chaouloff F. Serotonin_{1C,2} receptors and endurance performance [letter]. *Int J Sports Med* 1994; (15): 339
 102. Bailey S, Davis J. Response to letter to the editor by F. Chaouloff [letter]. *Int J Sports Med* 1994; (15): 340-1
 103. Westerink B, Justice J. Microdialysis compared with other in vivo release models. In: Robinson T, Justice J, editors. *Microdialysis in the neurosciences*. Amsterdam: Elsevier Science Publishers, 1991; 23-46
 104. Ungerstedt U. Introduction to intracerebral microdialysis. In: Robinson T, Justice J, editors. *Microdialysis in the neurosciences*. Amsterdam: Elsevier Science Publishers, 1991; 3-22
 105. Ungerstedt U, Hallström A. In vivo microdialysis, a new approach to the analysis of neurotransmitters in the brain. *Life Sci* 1987; 41: 861-4
 106. Benveniste H, Hüttemeier C. Microdialysis – theory and application. *Prog Neurobiol* 1990; 35: 195-215
 107. Kissinger P, Hart J, Adams R. Voltammetry in brain tissue: a new neurophysiological measurement. *Brain Res* 1973; 55: 209-13
 108. Cenci A, Kalen P, Mandel R, et al. Regional differences in the regulation of dopamine and noradrenaline release in medial frontal cortex, nucleus accumbens and caudate-putamen: a microdialysis study in the rat. *Brain Res* 1992; 581: 217-28
 109. Imperato A, Honore T, Jensen L. Dopamine release in the nucleus caudatus and the nucleus accumbens is under glutamatergic control through non-NMDA receptors: a study in freely moving rats. *Brain Res* 1990; 530: 223-8
 110. Meeusen R, Sarre S, De Meirleir K, et al. Microdialysis as a method to measure central catecholamines during exercise [abstract]. *Med Sci Sports Exerc* 1994; 26 (5): S23
 111. Pagliari R, Peyrin L, Milano S. Effect of submaximal physical exercise in norepinephrine release in the rat frontal cortex: a study with microdialysis. In: Louilot A, Durkin T, Spampinato U, et al., editors. *Monitoring molecules in neuroscience*. Gradignan: Publi Typ 1994; 342-3
 112. Hattori S, Li Q, Matsui N, et al. Treadmill running combined with microdialysis can evaluate motor deficit and improvement following dopaminergic grafts in 6-OHDA lesioned rats. *Res Neurol Neurosci* 1993; 6: 65-72
 113. Sabol K, Richard J, Freed C. In vivo dialysis measurements of dopamine and DOPAC in rats trained to turn on a circular treadmill. *Pharmacol Biochem Behav* 1990; 36: 21-8
 114. Meeusen R, Smolders I, Sarre S, et al. The effects of exercise on extracellular glutamate (GLU) and gamma-aminobutyric acid (GABA) in rat striatum, a microdialysis study [abstract]. *Med Sci Sports Exerc* 1995; 27 (5): S215
 115. Gerin C, Legrand A, Privat A. Study of 5-HT release with chronically implanted microdialysis probe in the ventral horn of the spinal cord of unrestrained rats during exercise on a treadmill. *J Neurosci Methods* 1994; 52: 129-41
 116. Guadalupe T, Perez-Rodriguez I, Gonzalez-Mora J. Involvement of nucleus accumbens dopamine in motor activity: a voltammetric study. In: Louilot A, Durkin T, Spampinato U, et al., editors. *Monitoring molecules in neuroscience*. Gradignan: Publi Typ 1994; 179-80
 117. Chaouloff F. Influence of physical exercise on 5-HT_{1A} receptor- and anxiety-related behaviours. *Neurosci Lett* 1994; 176: 226-30

Correspondence and reprints: Dr Romain Meeusen, Vrije Universiteit Brussel, Dept Human Physiology and Sportsmedicine, Laarbeeklaan 101, Brussels 1090, Belgium.